Fungal allergy in asthma-state of the art and research needs

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Abstract

Sensitization to fungi and long term or uncontrolled fungal infection are associated with poor control of asthma, the likelihood of more severe disease and complications such as bronchiectasis and chronic pulmonary aspergillosis. Modelling suggests that >6.5 million people have severe asthma with fungal sensitizations (SAFS), up to 50% of adult asthmatics attending secondary care have fungal sensitization, and an estimated 4.8 million adults have allergic bronchopulmonary aspergillosis (ABPA). There is much uncertainty about which fungi and fungal allergens are relevant to asthma, the natural history of sensitisation to fungi, if there is an exposure response relationship for fungal allergy, and the pathogenesis and frequency of exacerbations and complications. Genetic associations have been described but only weakly linked to phenotypes. The evidence base for most management strategies in ABPA, SAFS and related conditions is weak. Yet straightforward clinical practice guidelines for management are required. The role of environmental monitoring and optimal means of controlling disease to prevent disability and complications are not yet clear. In this paper we set out the key evidence supporting the role of fungal exposure, sensitisation and infection in asthmatics, what is understood about pathogenesis and natural history and identify the numerous areas for research studies.

Keywords

Aspergillus, Severe asthma, SAFS, ABPA, ABPM, Corticosteroid, Eosinophil, IgE, Itraconazole, Hypertonic saline

Introduction

Fungal exposure is a daily fact of human existence, which infrequently results in disease. Yet fungal allergy drives asthma severity in very large numbers of people affected by severe asthma. Available statements from different medical associations are unequivocal in declaring that fungi are sensitizers and exacerbate allergic asthma (American College of Occupational and Environmental Medicine [1], Institute of Medicine, American Academy of

Allergy, Asthma and Clinical Immunology [2] and American College of Medical Toxicology). Increasing rates of fungi-associated occupational asthma are also of concern. In contrast to allergy to other environmental agents, antifungal therapy is available, yet our knowledge of who, when and how to treat is in its infancy. As with other allergens, immunization could be helpful but has barely been addressed.

The European Academy of Allergy and Clinical Immunology (EAACI) Asthma Section decided to address this deficiency by forming a Task Force to address 'Fungal Allergy in Asthma'. The first product of the Task Force is a summary of what is and what is not known about this topic, written from the perspective of the progressive practicing clinician. It is focused on the lower respiratory tract, does not address cystic fibrosis (CF) in any depth and highlights research needs in the area. Other consensus reviews and statements relevant to this topic have described the status of allergic bronchopulmonary aspergillosis (ABPA) in cystic fibrosis [3], lower airways interaction with fungi and its clinical consequences [4] and the whole topic of ABPA [5], the last with recommendations for a new definition of ABPA.

Conceptual background

While fungal exposure is universal, sensitisation and disease are not. Very early life airborne contact with fungi is well demonstrated by studies with *Pneumocystis* serology and pneumonia in healthy children and those with cancer [6]. Broadly speaking, fungi can cause problems to the lung in two ways; either by acting as aeroallergens or as a pathogen causing infection. Some fungi can do both, often simultaneously. To cause infection in the lung the fungus has to be able to grow at body temperature and this property is restricted to a relatively narrow range of fungi, particularly yeasts and members of the *Aspergillus* and *Penicillium* genera. The commonest fungus causing lung infections is *Aspergillus fumigatus*, although other *Aspergillus* spp. are also implicated [7]. Fungal allergens, which can cause rhinitis and asthma, but rarely cause infection, include spores from the plant pathogens *Cladosporium* and *Alternaria* spp. A third potential cause of ill-health from fungi are volatile organic compounds and mycotoxins released by moulds such as *Stachybotrys* spp, which remains controversial and will not be discussed here in depth.

Sensitisation and allergy

Allergy is an inflammatory response caused by an environmentally delivered and often non-pathogenic agent and is caused by an exaggerated immune response rather than the pathogenic, pharmacological or toxic properties of the primary agent. As fungi are complex eukaryotes, all forms of allergic immune response [8] should be considered as potentially leading to fungal allergy, although the most well-recognised clinical responses, such as asthma and rhinitis caused by *Alternaria alternata*, are mediated in a straightforward immunoglobulin (Ig) E/TH$_2$ manner. The stipulation on including evidence of an inflammatory response in the definition of allergic disease is to distinguish *allergy* from *sensitisation*. Many people with elevated specific serum (s)IgE (or for that matter other intermediates of immune response) against a certain agent (sensitisation) do not develop symptoms when exposed to that agent. However, this is not a fixed difference as sensitisation can evelove into allergy depending on the level of exposure, co-factors present at the time of exposure and the age of person, with periods in their life when they develop symptoms and periods when they have sub-clinical disease or are in complete remission induced by immune tolerance.

Infection

Viable microorganisms including fungi can have a range of interactions with their human host. *Infection* refers to the presence of a microorganism, which leads directly to ill-health as a result of its pathogenic properties. *Colonisation* refers to any situation where a microorganism becomes established in a new environment and doesn't imply any particular relationship between the organism and the host. Both pathogenic and non-pathogenic organisms can act as commensals. The ability of microorganisms cause disease depends on their pathogenic potential, the number of organisms, the integrity of host defense, the strain of the organism and no doubt other factors, which are yet to be defined. If there is evidence of tissue dysfunction which is likely to be due to the presence of the microorganism then the term *infection* is preferable to colonisation.

Fungi can cause a number of different types of infection, separately from their ability to act as sensitisers. Contrasting patterns of fungi-host interactions are illustrated in Figure 1, with *A. fumigatus* remarkable as it can cause invasive infection in the immunocompromised, chronic pulmonary aspergillosis and *Aspergillus* bronchitis in non-immunocompromised individuals with underlying lung damage, and allergic disease of the upper and lower airways. In contrast, *Pneumocystis* is a pathogen of the immunocompromised, but not an allergenic fungus or sensitizer, which greatly differs from the primary skin pathogen *Trichopyton interdigitale* (and other species) which causes cutaneous infection in hundreds of millions of people, cannot grow in the lung, but is a common sensitising fungus associated with severe asthma.

Figure 1 Patterns of fungal interactions with humans, illustrating different host pathogen interactions, based on the host damage response framework [9]**.**

Asthma

This position paper concerns fungal allergy in *asthma*. However asthma itself is not a straightforward term and requires clarification. Asthma can be defined in two ways, descriptively and rather loosely as a multifactorial chronic inflammatory disease of the airways with an associated set of typical symptoms and evidence of variable airflow obstruction, or more strictly as a physiological abnormality in which there is documented rapid changes in the resistance to airflow and increased sensitivity of the airway smooth muscle to bronchoconstricting stimuli (airway hyperresponsiveness (AHR)) [10]. Bio-statistical approaches to measuring heterogeneity in asthma have suggested that asthma as broadly defined is comprised of a number of distinct endotypes, not all of which have obvious AHR or variable airflow obstruction [11,12], and improvement in symptoms of asthma are not always reflected in better physiological measurement of airflow. Many conditions can mimic asthma, or occur alongside it, confusing the clinical picture.

Current disease concepts, definitions and terminology

We have tabulated the current and proposed definitions of fungal allergy, sensitisation and related conditions in Table 1. ABPA was first described in 1952 [13], and the criteria for diagnosis were defined during the 1970s and 1980s [14-17]. It is recognised that occasionally ABPA can occur in the absence of asthma or CF, with other criteria positive. The cutoff for total serum IgE has not been rigorously addressed in asthma, although in CF in the UK a

cutoff of >180 IU/mL was 91% sensitive and 90% specific [18] in contrast to an Indian cohort of asthmatic patients with ABPA in whom a total serum IgE of 2346 IU/mL (sensitivity 87.5%, specificity 66.9%) appeared optimal [5]. The utility of the Patterson criteria have been prospectively evaluated and compared with recently introduced criteria by Agarwal [19]. Use of 6 (but not 5, 7 or 8) 'Patterson criteria' performed best in a cohort of 372 asthmatics in northern India [19]; 98% of the 56 patients with ABPA identified in this study had bronchiectasis, potentially affecting the performance of the definition. In addition, some clinicians use expectoration of brown coloured mucus plugs as a minor diagnostic criteria for ABPA [20], observed in 31 to 69% of patients [5]. These results raise questions about the optimal disease model and as a consequence the diagnostic criteria for ABPA (see Figure 2).

Table 1 Definitions

Entity	Current definition	Proposed definition	Comments
Fungal allergy	Immune-mediated inflammatory response to a fungus sometimes leading to tissue damage	Same, being inclusive of all allergic immunopathologies	Demonstrating and documenting 'Tissue damage' can sometimes be difficult.
Fungal sensitisation	Immune-mediated response to a fungus, without evidence of inflammation or tissue damage, usually documented by an elevated fungal-specific IgE.	Same	Tends to reflect specific-IgE response (or skin prick test result) only.
Fungal colonisation	None	1. One (or preferably two or more) respiratory sample (s) positive for a fungus by culture or PCR 2. No new major respiratory symptoms 3. No evidence of ABPA or other forms of aspergillosis 4. No overt immunocompromise 5. Negative fungal specific IgG in serum.	Such criteria may apply to other filamentous fungi, but not Candida. They need to be tested in prospective studies.
ABPA	1. Asthma (or CF) with deterioration of lung function 2. Elevated total serum IgE of >1000 ng/ml (>417 IU/ml) 3. Elevated *A. fumigatus* specific IgE and/or IgG antibodies 4. Immediate *Aspergillus* species skin test reactivity 5. Eosinophilia (>1,000/uL) 6. Presence of central (or proximal) bronchiectasis 7. Chest radiographic infiltrates 8. High attenuation mucus present.	1. Asthma (or CF) 2. *A. fumigatus* skin test positive or elevated *A. fumigatus* IgE levels 3. Elevated total serum IgE of >1000 IU/ml + 2 of the following: a. Positive *A. fumigatus* IgG or precipitating antibodies b. Radiographic opacities consistent with ABPA c. Eosinophil count >500 cells/uL	No definition addressed robustly with prospective study and combinations of diagnostic criteria.
Allergic bronchopulmonary mycosis (ABPM)	1. Asthma (or CF) with deterioration of lung function 2. Elevated total serum IgE of >1000 ng/ml (>417 IU/ml) 3. Elevated fungal specific IgE and/or IgG antibodies 4. Immediate fungal species skin test reactivity 5. Eosinophilia (>1,000/uL) 6. Presence of central (or proximal) bronchiectasis 7. Chest radiographic infiltrates	No new proposal, but similar to ABPA, with substitution of a different fungal specific tests.	Too rare to develop patient cohorts to formally validate a definition.
Severe asthma with fungal sensitisation (SAFS)	1. Severe asthma 2. Total IgE <1,000 IU/mL 3. Sensitisation to any fungus by skin prick test or sIgE		Severe asthma is a variable, usually treatment-based entity. Variable performance of different skin and sIgE test reagents, makes SAFS an imprecise entity until diagnostics improve.
Aspergillus bronchitis	None	1. Multiple respiratory sample positive for *Aspergillus* spp by culture or PCR 2. Major respiratory symptoms for >4 weeks 3. No evidence of ABPA or other forms of aspergillosis 4. No overt immunocompromise 5. Positive *Aspergillus* IgG or precipitins in serum.	Few patients reported. Some cases are caused by non-*fumigatus* species, and so serology criterion may be falsely negative.

Fungal allergy, fungal sensitisation and fungal colonisation may be clinically silent.

Figure 2 Two contrasting disease models (A and B), with common elements of risk and exposure, but different outcomes. Model A illustrates a linear relationship between disease and complications, whereas **model B** provides independent consequences of fungal exposure, which may or may not be associated. In both models, it is proposed that airway colonization/infection with fungus, and possibly fungal impaction from breathing airborne non-pathogenic fungi, and/or bacterial infection, continue to drive the inflammatory process.

There are some other difficulties with current diagnostic criteria. One of these is overlapping syndromes: separation of ABPA from Hyper IgE Syndrome (Job's syndrome) [21], separation of ABPA complicated by bronchiectasis and *Aspergillus* bronchitis [22], separation of ABPA from chronic pulmonary aspergillosis with a prominent TH_2 response, including raised serum total and sIgE [23], and Churg-Strauss syndrome in the absence of biopsy as common examples. While the presence of central (or proximal) bronchiectasis and/or chest radiographic infiltrates are usually cited as primary criteria of ABPA [4,24], a recent consensus group suggested that bronchiectasis should be regarded as a complication of ABPA and avoidable may be with earlier therapy [5].

It has been recognised that other fungi can induce a disease process similar to ABPA, normally called allergic bronchopulmonary mycosis (ABPM) [25]. Many different fungi can be implicated and the remarkable feature in all is a very high total and specific serum IgE to one or more non-*Aspergillus* fungi, and other findings usually found in ABPA. *Candida albicans* is the most commonly reported associated fungus with *Bipolaris* species, *Schizophyllum commune* and *Curvularia* species the next most common.

In 2006, the term severe asthma with fungal sensitisation (SAFS) was introduced [26] and subsequently shown to be responsive to antifungal therapy [27]. For consistency, the definition of severe asthma should be uniform, but is often not fully distinguished from dysfunctional breathing and tracheomalacia. In addition, current definitions of severe asthma generally reflect treatment intensity; the WHO identified 3 patterns: 1) untreated severe asthma, due to undiagnosed asthma or unavailability of therapy, 2) difficult-to-treat severe asthma (due to adherence issues, inappropriate or incorrect use of medicines, environmental triggers or co-morbidity), and 3) treatment-resistant severe asthma, including asthma for which control is not achieved despite the highest level of recommended treatment or asthma which is controlled only with the highest level of recommended treatment [28]. Asthma severity may also change with antifungal therapy, which was vividly illustrated by a report of antifungal therapy allowing one third of patients having their asthma severity downgraded [29]. The definition of SAFS includes people sensitized to purely allergenic fungi such as *Alternaria alternata* but sensitisation to thermotolerant fungi is not restricted to people with severe asthma, nor indeed to people with asthma, creating uncertainty with respect to the best disease model. Sensitization to *A. fumigatus* is more common in severe asthma but it is not uncommon to find patients who present with severe fixed airflow obstruction on a background of apparently mild to moderate asthma who are sensitized to *A. fumigatus*, suggesting the tissue damage associated with allergy to *A. fumigatus* is not restricted to severe disease. While ABPA was accepted as an endotype of asthma by another EAACI Task Force [12], SAFS was not and remains a pragmatic definition to enable an antifungal therapy trial. This issue needs resolution, possibly by genotyping patients with different forms of asthma [30], if genotypes allow such classifications, which they may not.

Fungal sensitisation is also found in allergic rhinosinusitis, without asthma or ABPA, and is a diagnostic criterion for allergic *Aspergillus* rhinosinusitis [31,32]. Recently over 10% of

patients with chronic obstructive airways disease (COPD) were found to be sensitized to *A. fumigatus* and this was associated with worse pulmonary function [33].

The role of airway colonization or infection by fungi, is probably significant, and is most notably related to *A. fumigatus*. About 60% of asthmatics sensitised to *A. fumigatus* have *A. fumigatus* growing in their sputum on a single sample (compared to ~5% of normal subjects) and this increases to 80% if multiple samples are obtained over time (AJW/CHP personal observation). Importantly, these patients have impaired lung function and increased rates of bronchiectasis compared to matched controls suggesting evidence of tissue damage [24]. Likewise those asthmatics sensitised to *A. fumigatus* have a much higher rate of bronchiectasis [34], and children sensitised to *Alternaria* are more likely to have persistent asthma in adulthood [35]. Our supposition is that fungal airway infection induces an ongoing allergic stimulus, as although most fungal conidia (spores) release some allergen(s), many more are produced after spore swelling and germination [36]; before germination the conidia are covered in hydrophobin proteins, which elicit no immune reaction [37]. Preformed allergen from hyphal fragments may elicit an immune response, even in the absence of local germination and growth [38], possibly explaining why increased environmental exposure to *Cladosporium* spp. or *Alternaria alternata*, for example are associated with worse asthma.

Another clinical entity, widely acknowledged in several southern countries, is "*Tricophyton Asthma*". There are multiple lines of evidence for sensitisation to *Tricophyton* proteins in asthma patients [39-42], where inhalation and/or dermal absorption of *Trichophyton* antigens are currently considered as possible routes of exposure [39,41]. Another study showed a high positive skin test rate in asthmatic patients with *Trichophyton* infection, confirming that the presence of cutaneous fungal infection is a crucial determinant of sensitivity to *Trichophyton* [42], regardless of the patients' atopic status [41]. A Japanese study showed that specific IgE response to *Trichophyton* doubles to 32.4% with increasing severity of asthma, suggesting an independent determinant of asthma severity [40]. Otherwise, there are few data about the relative frequency of *Trichophyton* sensitisation in asthma. One recent UK study found a frequency of 17% in polysensitised severe asthmatics; 10% were sensitised to *Trichophyton* as a single mould sensitisation, but only 2.5% were sensitized to *Trichophyton* alone [43].

What terminology can we use to capture the full spectrum of disease characterized by fungal sensitization/allergy, often associated with fungal airway infection? ABPA/M has a well-established but restrictive definition, leaving out the majority of patients sensitized to *A. fumigatus* and related moulds without evidence of lung damage. Perhaps ABPA, ABPM and SAFS are simply the severe end of the same spectrum of disease. On the other hand, sensitization to moulds is also seen in people with mild asthma and good lung function or allergic rhinitis where the mould may simply act as an allergen, although it is possible that over time these people will develop fixed airflow obstruction, bronchiectasis and even chronic pulmonary aspergillosis and parenchymal fibrosis. Ideally, definitions and inclusion criteria should be based on outcomes, for example treatment response to antifungal agents or risk of developing progressive lung damage. However, there are virtually no robust data to guide this approach. One simple option would be to adopt an inclusive approach and relax the criteria for ABPA/M to describe anyone with airway disease, evidence of allergy to thermotolerant fungi (as defined by a positive serum sIgE or SPT) and lung damage (fixed airflow obstruction, tree in bud shadowing on CT scan, bronchiectasis of any description, or lung fibrosis). In this model, people with airway disease and sensitisation to thermotolerant fungi, but no evidence of lung damage, would be regarded as at risk of developing future lung damage and kept under appropriate surveillance. If this model were adopted it would need to

be followed by large-scale trials of antifungal and/or immunomodulatory agents and large longitudinal cohort studies powered to relate the various immunological and clinical features of ABPA/M in its new definition to the risk of lung damage and the benefits of eradication of airway fungal infection.

Disease concept research priorities

- *Large longitudinal observational studies of people with airway disease including asthma with different severity and sensitization to* A. fumigatus *and other fungi powered to determine the relationship between development or progression of lung or airway damage and the clinical and immunological criteria currently used to define ABPA/M, SAFS and sensitisation.*
- *Large studies of effective anti-fungal agents in people with airway diseases including asthma of different severity and sensitization to* A. fumigatus *and other fungi powered to determine the relationship between treatment outcomes, the course of disease and the clinical and immunological criteria currently used to define ABPA/M and SAFS.*

Natural history and key findings

ABPA has a natural history that is both poorly characterized and difficult to predict, but better understood than other entities of fungal allergy in asthma. ABPA does not necessarily occur in the most severe cases of asthma. Sensitization to fungi is rare in childhood, although ABPA occurs occasionally in children [44-46], as does the recently described SAFS. Sensitisation to *Alternaria* in childhood (age 6 years) is associated with persistent asthma in early adulthood [35], but no other fungal allergens were tested. The natural history of fungal sensitisation in mild or moderate asthma is not currently described at all.

Patterson et al. described five non-sequential stages of ABPA (acute, remission, exacerbation, steroid-dependent asthma and fibrosis) based on clinical presentation, including recurrent and/or chronic persistent symptoms [47,48]. This staging, however, does not necessarily reflect the disease's progression. Although the disease may remit temporarily, it is more often a progressive disorder with recurrent, infrequent acute episodes that cause successive bronchial damage. Tissue damage tends to be greatest in the proximal airway region, where mycelial plugs and antigen production are localised, giving rise to granulomatous airway inflammation that results in the characteristic proximal bronchiectasis [49]. In the early stages of ABPA, the bronchial wall is infiltrated with mononuclear cells and eosinophils. If mucoid impaction and atelectasis occur, they may be followed by bronchiolitis obliterans, granulomatous bronchiolitis, and pulmonary fibrosis [50].

A large variety of imaging changes are found in ABPA, and may be related to the phase of the disease. Lung infiltrates, bronchiectasis and mucoid impaction are the most common pathologic findings seen on a chest radiograph with ABPA. Transient changes include areas of consolidation. The pathologic basis of consolidation is unclear: it may be related to airway obstruction or eosinophilia. Although such areas of consolidation are usually transient, some patients present with permanent consolidation, perhaps due to endobronchial obstruction. When consolidation clears, it often leaves residual bronchiectasis. The pulmonary findings of ABPA on high-resolution CT (HRCT) include centrilobular nodules, bronchiectasis (often with mucoid impaction), fibrosis and cavitation. In SAFS, only bronchiectasis is well recognised, although both airway and peripheral pleuroparenchymal fibrosis, a manifestation of chronic pulmonary aspergillosis, is also described [51].

ABPA has been classified either in terms of ABPA-S and ABPA-CB (depending on, respectively, the absence (ABPA-S) or presence of bronchiectasis (ABPA-CB)) or in terms of ABPA-S (mild), ABPA-CB (moderate) and ABPA-CB-ORF (other radiologic findings) [52-54]. The prevalence of bronchial dilatation in patients with severe or persistent asthma ranges from 17% to 30%, compared with 90% or more in ABPA [55]. It has been thought that patients with ABPA-S represent the earliest stage of the disorder, but prolonged follow up of this group showed no interval development of bronchiectasis, despite exacerbations [56]. In a study of 126 patients, the clinical, spirometric and immunologic findings were not significantly different when classified in terms of either ABPA-S and ABPA-CB or ABPA-S, ABPA-CB and ABPA-CB-ORF [57].

High-attenuation mucus (HAM) is a characteristic radiologic finding seen in patients with ABPA [57], and not in any other asthma entity. A multivariate analysis showed that severity of bronchiectasis and presence of hyperattenuating mucus impaction are predictive of ABPA relapse, and that severity of bronchiectasis was an independent predictor of failure to achieve long term remission [58]. In the same study, HAM was shown to be present in 18% of patients diagnosed with ABPA. Reasons why hyperattenuating mucus is associated with poorer outcomes remain unclear; high attenuated mucus may be a more inspissated type of mucus. Perhaps this subgroup of patients has more severe inflammation and/or specific genetic alterations that favour the formation of HAM [58]. More research is therefore required to investigate the exact reason for this association. It is not clear whether bronchiectasis and pulmonary fibrosis constitute sequels of mucoid impaction in ABPA. Mucoid impaction as an entity is not well understood, and may be associated with *Aspergillus* bronchitis (usually complicating bronchiectasis) and obstructing bronchial aspergillosis (in immunocompromised patients).

The relapse frequency in ABPA is usually low, and reduced further by corticosteroid and antifungal therapy in many patients. Whether significant external fungal exposures precipitate relapse is not clear, but possible. Viral and bacterial respiratory infections may contribute to relapse, or to associated tissue damage and development of complications. It may be that remission of ABPA and SAFS occurs in older age, but this is not studied.

Pulmonary cavitation and fibrosis are often severe complications of ABPA, and may occur in SAFS. In one series, cavitation and atelectasis were shown to develop in, respectively, 20% and 46% of patients with ABPA [59]. Cavitation was observed to occur in patients with a chronic infiltrate and was thought to represent necrosis within an eosinophilic pneumonia. Cavitation may be reversible, following resolution of infection. The development of chronic pumonary aspergillosis (CPA) with or without cavitation may complicate ABPA and aspergillomas have been reported in up to 7% of patients with ABPA. Aspergilloma usually occurs in patients with longstanding ABPA and extensive lung destruction, even though this is not always the case: rapid formation of aspergilloma in early ABPA has also been described (before bronchiectasis), implying that in some patients, it may be formed from direct parenchymal invasion [60]. In a recent study, ABPA was identified as an underlying condition in 18 (14.3%) and, more specifically, as the primary underlying condition in 15 (11.9%) out of 126 CPA cases, some with aspergillomas [51]. These findings support those from a study of aspergillomas where ABPA was identified as an underlying condition in 11.8% (10 out of 85) of cases reviewed [51]. The fact that ABPA is a relatively common underlying condition suggests that, it is not so much the presence of *Aspergillus* itself, as are the interaction of the lung's condition (bronchiectasis/fibrosis) and corticosteroid exposure (which is the treatment of choice for ABPA) that are the risk factors for development of CPA.

The radiological manifestations of early CPA in ABPA are not well delineated. Whether antifungal therapy arrests the progress of CPA complicating ABPA is not clear, but our anecdotal experience suggests it does. SAFS has also been associated with CPA, though infrequently [51].

Invasive aspergillosis complicating ABPA has also been rarely described, the first time in 1975 [61].

Natural history research priorities

- *Large longitudinal observational studies of people with ABPA and SAFS, including detailed imaging, from as early a stage as possible after recognition to ascertain the relative frequency and associations with the different complications.*
- *Detailed study of mucoid impaction and hyper-attenuating mucus.*
- *Genetic and functional studies comparing patients with and without complications of ABPA and SAFS, including those with mild and severe bronchiectasis and those with early and advanced CPA.*
- *A better understainding of co-morbidities and their impract on natural history, is also required.*

Epidemiology

Asthma prevalence and burden of fungal allergy in asthma

The Global Burden of Disease 2010 project estimated that asthma was ranked 43rd globally for years life lost [62], although 15th and 20th in Oceania and Eastern Europe respectively; 15th globally in terms of prevalence with an estimated 334 million people affected; 14th in terms of years lived with disability [63]; and 28th in terms of disability-adjusted life years [64]. An estimation of the adult burden of asthma using an alternative modeling approach based on the global initiative for asthma (GINA) yielded a global burden of 197 millions [65]. Utilising the 5 estimates of ABPA period prevalence (0.7-3.5%, median 2.5%) among referred patients to secondary care from South Africa, Ireland, Saudi Arabia, New Zealand and China there are 4.8 million adults with ABPA complicating asthma [65]. This estimate could be an overestimate because only a proportion of cases is referred to secondary care, or an underestimate because milder cases are not investigated in the community or referred, or were investigated previously and discharged back to the community.

Global or national SAFS prevalence estimates have not been made. The more severe asthma is, the higher is the frequency of *Aspergillus* and fungal sensitisation. Assuming that the worse 10% of adult asthmatics are severe and that fungal sensitisation has a minimum frequency of 33% in these people, about 6.5 million would be expected to have SAFS, although there is some crossover with ABPA patients who also have severe asthma [65]. It is not known how much regional or ethnic variation there is in SAFS prevalence.

Environmental exposure

Many recent studies describe mould-related respiratory effects in asthma [66-68]. Studies in environmental medicine have established a clear causal relationship in mould-induced asthma. Furthermore, these studies indicate that prevalence of adulthood asthma induced by

moulds is increasing. The fraction of adulthood asthma incidence attributable to work is about 30% [69]. Adult-onset asthma is strongly associated with the level of fungal exposure [70]. Quite recently, moulds became the leading causative factor for occupational asthma (OA); notably in Finland. All *de novo* diagnosed cases of occupational asthma in Finland are obligatorily reported to the National Register, and by 2002 moulds become the most important causative agent (18% of cases in 1998-2002) [71]. Importantly, diagnosis of mould-induced asthma was confirmed by the allergen-specific challenge tests. The people affected are mostly white-collar employees, a population not generally considered as a risk group for OA. The same increasing trends have been shown in UK [72]. One of the most well established causes of occupational asthma associated with fungal allergens is alpha amylase from *Aspergillus oryzae* which is used as a additive in bakeries and can lead to 'bakers asthma' [73]. Alpha amylase has also been implicated in case reports in causing allergic reactions to white bread [74].

Outdoor

The data on the association between total fungal spore counts, spores grouped by their division or individual spore *taxa* and hospitalizations or visits to emergency departments due to asthma exacerbations are inconsistent. Recently, the relative abundances of fungal genera were evaluated using pyrosequencing approach in outdoor air samples from an urban US area and estimated relative abundance of fungi recognised to be allergenic from 2.8% to 10.7% of total fungal taxa [75]. Several studies did not find associations of outdoor fungal spore counts and visits to emergency departments or hospital admissions due to asthma exacerbations in adults [76,77], although both studies did find these associations in children. Fungal sensitisation was not investigated in these studies. In contrast, some studies have shown that sensitization to fungi *per se* is a risk factor for admission to an intensive care unit with an acute attack of asthma [78] and respiratory arrest [79]. Subjects who had been admitted to the ICU with severe life-threatening asthma were significantly more likely to have one or more positive skin tests for fungal dry weather spores (*Alternaria alternaria, Cladosporium cladosporoides, Helminthosporium maydis, Epicoccum nigrum*). O'Halloren et al. found that subjects who had a respiratory arrest during the *Alternaria* aeroallergen season were more likely to have a positive skin test to *Alternaria* than other subjects with asthma [79]ss. Delfino and coworkers found associations between fungal spore concentrations, particularly *Alternaria* and *Helminthosporium*, and increased asthma symptom scores, decreased evening PEF and β-agonist inhaler use in adult asthmatics sensitized to fungi [80]. Additionally sensitisation to *A. alternata* in asthmatic subjects with grass pollen sensitivity was found to predict susceptibility to thunderstorm-associated asthma [81]. Children living in rural environments are less likely to have asthma and fungal sensitisation [82], whereas adults are more likely to have ABPA, at least in India (47% vs. 66%, p = 0.007), which may or may not reflect airborne exposure [83]. It is likely however that these differences are not simply about indoor or outdoor air exposures, as the mould exposures in indoor air were higher in rural environments (in France) [84], and non-allergic mechanisms of asthma exacerbation were suggested.

Indoor

Moulds are commonly present in houses; a study evaluating the presence of different mould species in more than 1000 houses in Northeast USA revealed that *Cladosporium* spp is found in 85%, *Penicillium*, *Aspergillus* and *Alternaria* respectively in 75%, 50% and 28% of homes [85]. Reported mould exposure and recent water damage is associated with asthma symptoms

and bronchial hyperresponsiveness in adult asthmatics [86]. This observed effect was consistent among 38 study centers in Europe and more prominent in subjects sensitized to *Cladosporium*. Sensitization to moulds is more prevalent in subjects leaving in damp dwellings compared to subjects living in dwellings without any sign of building dampness (9.3% vs 3.9% [87]). Furthermore, living in damp dwellings *per se* is associated with current asthma, lower FEV1 [88] and higher PEF variability [87]. Subjects sensitized to moulds, particularly to *Cladosporium* and *Alternaria* are at significantly higher risk of current asthma if they reported mould exposure [86,87,89]. Prevalence of current symptomatic asthma and use of asthma medications increase significantly with higher *Alternaria alternata* allergen levels indoor [90]. One study has shown that severity of asthma correlates significantly with measures of total dampness and mould growth in the dwelling [91]. However, numerous other studies were not able to confirm these findings. Wood and coworkers [92] did not find association between indoor mould and positive skin tests for common moulds. Several studies have shown no association between visible indoor moulds or airborne moulds and bronchial hyperresponsiveness [93,94].

Epidemiology research priorities

- *Detailed and population based studies of the prevalence and annual incidence of SAFS, fungal sensitisation and ABPA/M, in adults and children.*
- *Improved and more geographically diverse exposure studies relating the nature of exposures to asthma and ABPA severity.*

There is accumulating evidence that mould exposure is associated with asthma, however no clear exposure-effect relations have been established. The only population-based incident case–control study providing such evidence has shown increased risk of asthma in relation to sIgE antibodies to *A. fumigatus* and *C. herbarum* [95]. Interestingly, the majority of studies do not provide evidence that increased prevalence of asthma associated with often assumed exposure to moulds is caused by the hypersensitivity to fungi. Furthermore, no consistent associations were found for the objective measures of asthma severity and control. This is in part because of inconsistency and inadequate validation of the measures used to evaluate exposure and health effects. Therefore, it currently remains impossible to set evidence-based guidelines for avoidance of fungi. Perhaps colonization or infection of the airways in affected people overshadows external fungal exposure, although a recent study found *A. fumigatus* isolation from sputum to be associated with elevated airborne levels in homes of patients with asthma [96].

Pathogenesis

The pathogenesis of ABPA has been extensively studied, unlike fungal sensitisation and associated asthma. Although there is a correlation between the degree of exposure to *Aspergillus* spores and the development of ABPA, many findings suggest that ABPA is primarily the result of abnormal host immune response to *Aspergillus* antigens. Given that high incidence rates of ABPA have been reported in a number of families, such an abnormal host response is probably influenced by genetic factors [97,98]. While factors that allow growth of *Aspergillus* in the airways of ABPA patients remain unclear, bronchial mucus abnormalities have been implicated. For example, for unknown reasons, the mucus in ABPA can become extremely viscid and difficult to remove [99]. Clearly, complex interactions between host susceptibility and fungal development in the local pulmonary environment occur and need to be deciphered.

Lung microbiota

The application of sequence-based identification and metagenomics to studies of the human microbiome has revealed a plethora of organisms that can inhabit the human body including new or as yet unknown pathogens [100-102]. Most published studies have focused on bacterial communities, but fungi also form an important component of the microbiome [103]. The internal transcribed spacer regions (ITS1 and ITS2) of the nuclear ribosomal operon are the most popular loci used to identify and discriminate between fungal species and reference databases have been developed to allow comparison and identification of amplicons [104,105]. These regions have been the target of the few published lung fungal microbiota studies [104,106,107]. Unfortunately none have addressed ABPA, although a massive number of fungal species have been found in pooled sputum samples from asthmatics [108]. One area of uncertainty is the pathology of fungal growth within the lungs. Although in florid cases hyphae can be seen in mucus and there are old descriptions of bronchocentric granulomatosis (a distinctive immunopathology of allergic fungal disease), the extent and location of fungi in ABPA and SAFS affected lungs have not been well described.

Whilst fungal components do not need to be viable to elicit an allergic reaction, infection can only be caused by viable fungal cells. Conventional DNA-based methods for microbial identification are unable to differentiate between viable and nonviable cells, often resulting in an overestimation of microbial targets, which could be a major limitation. One approach that may circumvent this is viable-PCR (v-PCR), which utilises propidium monoazide (PMA) to detect only viable cells [109]. V-PCR has been applied to a number of microorganisms, including fungi, and has been used in conjunction with a number of DNA-based techniques including next generation sequencing. In sputum samples from individuals with cystic fibrosis, comparison of PMA treated and untreated clinical samples indicated that dead bacterial cells significantly bias untreated profiles [110].

Although high-throughput technologies have facilitated the identification and characterization of the human microbiome at mucosal sites including lungs, the mechanisms by which the commensal flora might influence lung immunity and their role in the development of allergic inflammation are not well characterized. Lung microbiota analysis is a new translational research area, offering the potential to redefine the processes that drives the progression of respiratory disease in asthma. Such studies will clearly impact our patient management, changing current paradigms of allergy/asthma and therapy.

Host-pathogen interactions

Neglect or react?

Every day we breath 20,000 times, thereby inhaling ~10,000 liters of air, which during the fungal season can often contain >50,000 fungal spores per cubic meter of air per day [111]. The size of these spores (approximately 2-50 μm) enables the smallest to reach the distal airways. For immune homeostasis it is critical to prevent excess inflammation and tissue damage in response to harmless species [112,113]. In response to the inhaled fungi, the pulmonary immune system has to decide whether to:

(i)   tolerate the presence of these fungi or
(ii)   to initiate an anti-fungal host defense program.

This decision and the resulting downstream signaling are critical for the integrity and maintenance of pulmonary host defense. These immunological 'neglect' or 'react' checkpoints are of key relevance for inhaled opportunistic fungi, in particular the filamentous saprophytic *A. fumigatus* or commensals, such as *Candida* species. These fungi can both colonize and infect the airways not only upon systemic immunosuppression, but also in pulmonary diseases where the local mucosal host defense is compromised, such as allergic asthma and cystic fibrosis [4,114-117]. Fungal colonization is the necessary antecedent event for sensitisation in the pathogenesis of ABPA, but it is unclear why only some individuals become sensitised. The factors determining the timing of sensitisation are not known and likely relate to a concurrent viral exposure and/or genetic predisposition (see Figure 2).

React

The pulmonary immune barrier function is mainly achieved by the lining epithelial cells, alveolar macrophages and pulmonary dendritic cells (DCs). These innate immune cells, are the first sentinels to recognize and react to inhaled fungi. Upon contact with fungi, airway epithelial cells produce large amounts of cytokines and chemokines, such as the prototypic interleukin 8 (CXCL8), which recruit leukocytes to the pulmonary site of infection to enhance antifungal host defense activities. The outcome of this initial interaction orchestrates the subsequent immune response. Macrophages phagocytose fungi, neutrophils phagocytose and/or release their granule contents and form DNA neutrophil extracellular traps (NETs) to entangle, immobilize and kill the fungus, a process involving neutrophil-derived calprotectin [118,119]. Even NK cells were recently found to potently fight *Aspergillus fumigatus* [120]. However, some fungi are capable of evading these host defense mechanisms, for instance *Candida*, by affecting phagosome maturation and phagosomal escape by formation of hyphae that destroy the vesicle and subsequently the entire host cell [121,122]. Neutrophils play the prime role in controlling *A. fumigatus* invasion and are essential in restraining tissue invasion and fungal spreading from the airways into the bloodstream [123,124]. In case of the allergic airway response to *Aspergillus*, the TH_2-associated chemokine CCL17/TARC plays a key role in both murine and human fungal asthma [125,126]. CCL17 binds to CCR4 and thereby recruits TH_2 cells into the airways that, in turn, drive IgE production and mast cell degranulation and feed the pro-allergic/-asthmatic airway response. Furthermore, CCL17 has been shown to impair macrophage killing of *Aspergillus* [125]. Since CCR4 is also expressed on Treg populations, these immunosuppressive cells may additionally favor survival of the fungus and a shift towards a TH_2 response. Clinically, CCL17 may serve as a serum biomarker in patients to differentiate sensitization to *Aspergillus* from clinically active ABPA flares, especially when quantified longitudinally, as supported by two studies [127,128]; cutoffs are not established. Inspired by findings in animal models of ABPA, anti-CCL17 antibodies are currently being developed (Hogaboam CM, *personal communication*).

Among cells of adaptive immune system, particularly IL-17-secreting T-helper cells (Th17) play a key role in anti-fungal host defense [129,130]. Autosomal recessive IL-17RA and autosomal dominant IL-17F deficiencies have been reported in chronic mucocutaneous candidiasis (CMC) patients [131]. A recent study further shows that IL-17A itself is able to bind fungal cells, thereby causing nutrient starvation conditions [132]. The Th17 response works in concert with IL-22, IL-23 and Th1/Tregs responses to control fungal dissemination [133]. On the other hand, dysregulation of these pathways can drive exaggerated inflammatory responses/hyperinflammation as demonstrated in chronic granulomatous disease (CGD) in mice [134]. These studies also show that the indoleamine 2,3-dioxygenase (IDO), a key enzyme of tryptophan metabolism, is involved in CGD-associated inflammation

and tryptophan catabolites (kynurenines) could be therapeutically useful to regulate and dampen hyperinflammation [135,136] although the regulation of this pathway in human CGD is different [137-140].

How are fungi recognized?

Similar to other microbes, fungi are recognized through conserved pattern recognition receptors (PRRs) of the innate immune system. Two families of PRRs are essential in fungal recognition:

- C-type lectin receptors (CLRs) and
- Toll-like receptors (TLRs) [118].

Prototypic CLRs include dectin-1, complement receptors, and macrophage mannose receptor. Less well-described include DC-SIGN, dectin-2 (recognizing house-dust mite), DCL-1, Mincle and CLEC5a. These CLR receptors have in common that they are coupled to Syk, which enables signaling via CARD9 and subsequent NFκB downstream activation [141]. Activation of these receptors by whole fungi through a proposed 'phagocytic synapse' [142] or fungal components (prototypically ß-glucans) elicit anti-fungal effector activities, that are (opsono)phagocytosis, cytokine production and respiratory burst. A recent study highlights that the interaction of *Aspergillus* conidia with the key CLR dectin-1 in the lung mainly takes places intracellularly in acidified phagolysosomes, following the phagocytosis of fungi by leukocytes [143]. The pathophysiological and clinical disease relevance of these receptors is corroborated by the finding that genetic mutations in distinct CLRs, namely dectin-1 [144] and CARD9 [145], cause CMC and other fungal infections. Beyond these pathways, STAT1 mutations have also been associated with CMC [146]. While genetic inheritance for susceptibility to *Candida* has been established, the molecular genetic basis for *Aspergillus* infections in humans remains to be defined.

In addition to CLRs, TLRs also recognize fungal patterns. The MyD88-coupled TLRs TLR1, TLR2, TLR4, TLR6 and TLR9 have been reported to contribute to fungal recognition in different experimental systems, with TLR2, TLR4 and TLR6 probably being the most important ones [147-150]. Cellular studies are supported by genetic association studies showing that one TLR4 polymorphism is weakly linked to susceptibility for invasive aspergillosis following haematopoetic stem-cell transplantation [151,152]. Studies with genetic knock-out mice mainly support the role of TLR2 and TLR4 in fungal recognition, but the specific receptor utilized depends on the phenotype of the fungus (stage-specific recognition, conidial *vs*. hyphal) and on the host genetic background (Brown GD, *personal communication*).

Besides cellular receptors, fungal patterns are recognized both by soluble innate immune receptors. Collectins, such as surfactant protein A (SP-A), SP-D, galectin-3, pentraxin-3, mannose binding lectin (MBL), C-reactive protein, complement, ficolins and others are *in vivo* the first proteins to bind fungi and facilitate the uptake and clearance by surrounding phagocytes. A recent study shows that opsonisation with surfactant components in the lung limits inflammation by attenuating host-fungi interactions, a mechanism relevant for zymosan but not for *Aspergillus* species [143], underscoring the complexity of host-fungus interactions in the lung.

Pathogenesis research priorities

- *Define the lower respiratory tract microbiome in ABPA, SAFS, ABPM and in asthmatics sensitised to different fungi, combined with v-PCR and other measures of microbial viability.*
- *Study specific fungal factors and host components that are worth targeting in patients, in order to (i) improve clearance of the pathogen and to (ii) limit hyperinflammation-mediated tissue damage.*
- *The mechanisms of airway and lung damage in ABPA and SAFS. The impact of treatments, including corticosteroids, on those mechanisms.*
- *Studies of the immunological response to fungi to determine the relationship between TH_2 allergic responses and Th1/Th17 infection related responses.*

Genetic associations

Several small studies have identified certain genetic factors linking the risk of ABPA with individual mutations. Multiple associations are to be expected given a family history in 5% of those with ABPA [98]. The genes implicated to date include IL-4Rα [153,154], IL-10 [155], surfactant A2 (SPA2) [156], TLR9 [30], CFTR [157-159], and HLA DR2/DR5 polymorphisms [160]. Some of these associations are not strong, and some may be related to disease progression or complications, rather than being risk alleles *per se*. All studies to date are relatively small. No genetic associations have been described with SAFS or other fungal allergic conditions.

Genetic association/risk research priorities

- *Expand confirm and evaluate the number of genetic associations with ABPA and other fungal allergic conditions, compared with asthmatic patients without these fungal disorders.*
- *Define genetic pathways that regulate the susceptibility towards and damage response pathways in pulmonary fungal infection. Genetic and functional studies comparing patients with and without complications of ABPA and SAFS, including those with mild and severe bronchiectasis and those with early and advanced CPA*
- *Risk associations in patients with and without disease could be direct association with disease or indirectly related to complications such as bronchiectasis or worse pulmonary function. Thus association studies should include well documented complication rates and functional status over time.*

Diagnosis and screening

A definitive diagnosis of 1the following 8 features to be present as follows: asthma; immediate *Aspergillus* skin prick test positivity (sensitivity and specificity: 94.7%, 79.7%); IgE levels >1,000 IU/mL (97.1%, 37.7%); positive *A. fumigatus* specific IgE (no value specified or studied) (100%, 69.3%); *Aspergillus* precipitins detectable (42.7%, 97.1%); eosinophil count >1,000 cells/uL (29.5%, 93.1%); transient or fixed chest radiographic opacities (36.1%, 92.5%); (central) bronchiectasis (91.9%, 80.9%); and high-attenuation mucus (39.7%, 100%) [5] (Table 2). So a key question is how these criteria should be applied in the routine clinical setting. Which are most useful as screening criteria to identify all those with ABPA? Further, can the same screening test(s) be used to identify patients with SAFS,

also deserving of a trial of antifungal therapy? These questions have recently been addressed by an International Society for Human and Animal Mycology (ISHAM) working group report which proposed screening with a blood *Aspergillus* specific IgE assay, and a simpler definition of ABPA (see Table 1) [5]. Overall, no single test has both good sensitivity and specificity; hence multiple tests should be utilized for confirmation of ABPA. The diagnosis algorithm proposed by the ISHAM Working Group is to perform an *Aspergillus* skin test and/or *A. fumigatus* specific IgE levels (the latter one being more sensitive) and if either is positive other tests for ABPA (CT scan, IgG specific to *A. fumigatus*, serum precipitins to *A. fumigatus* and total eosinophil count) should then be performed to establish the diagnosis of ABPA. What arguments are in favor of this approach, or another? Performance variability in the screening tests used to identify ABPA, SAFS, and those sensitised to fungi are pivotal to diagnosis, estimates of prevalence and understanding pathogenesis.

Table 2 Overview of screening tests applied to diagnosis of fungi-associated conditions

Test	Strengths	Weaknesses	Comment
Skin prick test (SPT)	Simple to perform. Rapid results. Good tolerability. Inexpensive. High negative predictive value (95%)	Accuracy and reliability dependent on quality of fungal extracts. Variability between different batches of test. Misses low sensitivity responses. Systemic and topical antihistamines may suppress weal and flare reaction. Presence of IgE without clinical symptoms.	Test should be performed with standardised allergen solution, if possible. Best used in conjunction with sIGE due to discordance in results.
Intradermal tests	More sensitive than SPT.	Higher rate of false positives than SPT.	Rarely used.
IgE (ImmunoCAP)[a]	Completely safe. Not influenced by concurrent drug treatment.	Results not immediately available. Testing more expensive than SPT. Presence of IgE without clinical symptoms.	Best used in conjunction with SPT due to discordance in results.
Complete blood count	Automated test.	Many different conditions result in increases or decreases in cell populations. Lack of correlation between peripheral blood eosinophil levels and lung function / immunological parameters.	Performed to assess peripheral blood eosinophil levels.
Fungal culture of sputum	Simple to perform. Inexpensive.	Lack of standardisation in most countries. UK standard insensitive.	Actively growing culture needed for strain-typing or anti-fungal sensitivity
Fungal PCR of sputum	More sensitive than culture.	May have high false-positives. Requires specialised equipment, although most labs have PCR machines.	Commercial tests available but no accepted standard for positivity.

a = often referred to as RAST tests, although the original radioallergoabsorbent test has been superceded by the ImmunoCAP or simpler ELISA tests.

The skin-prick test (SPT) is a simple diagnostic tool that can be useful for screening for ABPA [161], but it is not without limitations. The accuracy and reliability of the SPT and other *in vivo* and *in vitro* assays is highly dependent on the quality of the fungal extracts used. The quality can vary dramatically between commercial suppliers, which can be caused by inconsistencies in the preparation of fungal extracts; for example, using either fungal mycelia or spores for production. Inter-strain variability leads to extracts with altered protein composition, resulting in poorly standardised allergy testing solutions [162]. The majority of SPT positive individuals are also positive by specific IgE, giving the SPT a high negative predictive value (95%); however a significant proportion of individuals with positive IgE tests are SPT negative [4]. Intradermal tests are more sensitive than SPT [163]; however, these are rarely performed and carry a higher rate of false positives. *In vitro* measurement of multiple specific IgE antibodies may be more costly than SPTs. The CAP system for specific serum IgE testing has higher sensitivity than RAST tests with comparable specificity; and a recent study has suggested that both SPTs and specific IgE measurement by the (ImmunoCAP) system should be used in diagnoses of fungal allergy, due to discordance in test results in around a quarter of severe asthma patients [43]. Specific recombinant allergen testing may play a greater role in diagnosing ABPA in the future. Specific *A. fumigatus* allergens have been evaluated for their diagnostic performance in serologic studies in

asthmatic patients [164], and data suggests they may be useful in discriminating between ABPA (Asp f 2, Asp f 4 and Asp f 6) and fungal allergy (Asp f 1 and Asp f 3) [165,166]. Aspergillus precipitating antibody is found in <50% of ABPA patients and fewer SAFS patients, and is therefore a poor screening tool. Likewise radiology is insensitive and non-specific enough in ABPA and wholly unsatisfactory for screening for SAFS, in which radiology is often normal or nearly so.

Classic ABPA cases in the absence of oral corticosteroids have peripheral blood eosinophilia [167]; however, this feature may be only present at the time of exacerbation or during the acute phase of the disease [168]. A peripheral blood eosinophil count of above 1,000 cells/ul is often cited as a secondary diagnostic criterion and whilst it is still considered to be suggestive of ABPA [4] the utility of it as a diagnostic tool has been questioned [20,168], as most patients with ABPA have fewer eosinophils than this on presentation. One study to address this found no correlation between peripheral blood eosinophil levels and lung function or other immunological parameters; and whilst prevalence of central bronchiectasis was higher in patients with higher eosinophil counts, the severity of bronchiectasis did not correlate with the degree of peripheral eosinophilia [20]. Consensus of 500 cells/ul (at presentation or previously) was reached by the ISHAM working group, and needs prospective validation.

Culture of *A. fumigatus* from sputum is supportive but not diagnostic of ABPA, as the fungus can also be cultured from patients with other pulmonary diseases [169]. Sputum samples from individuals with asthma are usually culture negative [24]. The standard approach used to process sputum for mycological investigations in the UK has been shown to potentially underestimate fungal prevalence in the airway [170], and a recent multi-centre study has highlighted the variability in culture results that arise from different mycological methods being employed [171]. As such a number of groups have questioned the performance of the current UK respiratory culture methods for fungi and have called out for an improved, standardised approach [170-173]. PCR has been proposed as an alternative or additional approach to detecting *Aspergillus* species in sputum and has been shown to be more sensitive than culture in ABPA but needs to be interpreted with other clinical and laboratory features [174]. DNA-derived signals can originate from nonviable fungal cells [175,176] and therefore there is the potential for a positive PCR signal to derive from dead fungal material, particularly in the context of a patient who has been treated with antifungals.

Diagnosis and screening research priorities

- *Develop more standardized skin test and blood test reagents for detecting fungal sensitization, with validation in different populations with other potential causes of elevated IgE, such as chronic helminth infection.*
- *Assess the performance of better standardized tests for detecting ABPA and SAFS in asthmatics seen in routine clinical practice.*
- *Compare the performance of simpler combinations of tests to diagnose ABPA and SAFS, not including complications as a component of the diagnostic definition.*

To summarise, the current tests used to determine fungal sensitisation are not without limitations, and the discordance between test results must be taken into consideration. The use of specific recombinant allergens to discriminate between ABPA and fungal allergy has shown great potential and may be of importance in the future. The detection of *A. fumigatus* and other fungi from clinical specimens such as sputum is highly dependent upon the methodology used, and a more sensitive approach that is standardised and universally adopted is a definite need.

Clinical management

There are 5 aspects to the management of fungal allergy in asthma. These are:

1. Avoidance of fungi
2. Control of the inflammatory process
3. Improvement of airway air flow through reduction of mucus and obstruction
4. Reduction of fungal burden
5. Control of bacterial infection

Decreasing indoor exposure to moulds

Allergen avoidance remains the cornerstone in the management of any allergic condition. Several interventional studies employing remediation of damp buildings [177], removing visible mould [178], removing water damaged materials [179-181], applying fungicides [178], making alterations to heating [182], improving ventilation [178] and fixing buildings construction to prevent further water leakage [183] have been conducted. The interventions were conducted either at home [178,182,183] or in the working place [179-181,184,185].

Studies performed in occupational settings have shown that employees more commonly report improvement of symptoms while being away from work [179] and indicated a reduction of respiratory symptoms including cough, shortness of breath, wheezing [185], stuffy or runny nose, eye, throat, skin, and headache symptoms [179] after evacuation/relocation from the infected building [179,185]. In active intervention studies patients in remediation groups reported significant reduction of bronchitis [180], improvement in wheeze affected activities, perceived reduction in asthma medications (both relievers and controllers), symptoms of rhinitis [178] including allergic rhinitis [180], conjunctivitis [180,184], rhinoconjunctivitis [178], complaints on irritated eyes or nose and skin rash [181]. When remediation is compared to no remediation, a significant decrease in reported wheezing [178,183], need for asthma relievers and controllers [178] and trend toward fewer hospital admissions for respiratory conditions [183] in those patients whose

homes or work places had remediation undertaken. However, other studies failed to show a beneficial effect of intervention [182,184,186]. Inconsistent effects of remediation on lung function are also noted [178,180,181].

Farm environments could represent an occupational risk, as reported from India [83]. Specifically poultry farms may represent a local major source of fungi [187,188] and poultry workers have a higher rate of asthma [189,190] and mould IgG antibodies [189]. Protection of these workers may be important, but is not studied.

Numerous caveats accompany these interventional studies and they have several limitations. Interventions are applied indirectly to a particular participant as it the building is usually that receives attention. Moreover, usually it was impossible to determine a separate effect of mould reduction from interventions themselves such as increasing indoor temperature toward normal level could influence health. Some 'common sense' avoidance measures are usually recommended to affected patients such as avoidance of all composting, changing pillows regularly, avoiding building works, especially of old buildings, cellars and loft spaces. The evidence for these avoidance measures is limited [191]. Currently, additional studies are required to establish evidence-based guidelines on mould avoidance.

Control of the inflammatory process

Inhaled corticosteroids have transformed the management of asthma. Regular use in symptomatic asthmatics reduces mortality, but their impact in those with minimal or intermittent symptoms is not clear and they may be detrimental. One small prospective multicentre randomized controlled trial (RCT) in ABPA comparing beclomethasone and placebo showed no overt benefit and an increase in the frequency of *Aspergillus* isolation from sputum in the active group [192]. Despite beclomethasone, seven patients had clinical exacerbations as well as more radiological exacerbations than placebo. A 23 patient study of ABPA patients without bronchiectasis treated with inhaled corticosteroids showed subjective improvement but without complete control of asthma and median IgE levels increased [193]. The impact of inhaled steroids on ABPA (as opposed to asthma) is unclear and may only be necessary to improve asthma control. Inhaled corticosteroids are insufficient alone in controlling ABPA or fungal colonization, and could be detrimental in the latter, but this is unstudied.

There are no RCTs trials of systemic corticosteroids in ABPA although they are generally accepted as appropriate therapy for acute exacerbations. However few data are available to guide either dose or duration of corticosteroids. Lower doses of corticosteroids without antifungal therapy (0.5 mg/kg/day for 1-2 weeks, then on alternate days for 6-8 weeks then taper and discontinue) is associated with 45% more relapse and subsequent oral corticosteroid dependence [167]. A higher dosage of corticosteroid for longer (0.75 mg/kg for 6 weeks, 0.5 mg/kg for 6 weeks, then tapered by 5 mg every 6 weeks to continue for a total at least 6 weeks) was associated with a higher remission rate and a lower prevalence of corticosteroid-dependent ABPA (13.5%) [57]. Parenteral steroids may be required for ABPA unresponsive to oral steroids and antifungals, and may down-regulate steroid receptors [194]. Whether the benefit of short-term control with oral corticosteroids correlates with longer-term outcome has not been assessed. A RCT on the efficacy and safety of two different glucocorticoid dose regimens in ABPA (clinical trials.gov; NCT00974766) has been completed and results awaited.

Omalizumab has an established place in the management of some patients with severe asthma. Omalizumab reduces symptoms, exacerbations, asthma hospitalizations and has a steroid-sparing effect. While a recent study indicates that omalizumab is associated with improvements in outcomes in patients with uncontrolled persistent allergic asthma [195], the very high IgE in ABPA makes the use of omaluzimab difficult because of cost and administration limitations (volume). Whether those patients with SAFS should be first trialed on antifungal therapy or omaluzimab or given both is not known.

Macrolide therapy (especially azithromycin) has an anti-inflammatory role in some patients to decrease cough and sputum production. This unexpected role was discovered following the improvements noticed in patients with diffuse panbronchiolitis mainly found in people from East Asia [196]. It has been used in those with bronchiectasis, especially those with frequent exacerbations [197], as well as for other pulmonary diseases [198]. It has not been studied in ABPA and in particular the relative efficacy of macrolide versus itraconazole, in patients with and without bronchiectasis is lacking.

Improvement of airway air flow through reduction of mucus and obstruction

Nebulized hypertonic saline (6-7%, 4-5 mL) can be used to reduce the viscosity of sputum to ease expectoration of mucus plugs encountered in ABPA [199]. As hypertonic saline can also induce bronchoconstriction [200], caution is required with its initial administration especially in those with a low FEV_1. Its role in ABPA has not been actively addressed.

Mucus plugging of the airway is a characteristic feature of ABPA. Mucus is usually hypoattenuated, but may be hyperattenuated on CT scan. The removal of mucus plugs will generally improve clinical status and lung function. Indeed mucoid impaction is one uncommon presentation of ABPA. A course of oral steroids can sometimes relieve obstruction. The role of either nebulised hypertonic saline or N-acetyl cysteine in contributing to mucus plug removal is not studied. Therapeutic bronchoscopy (flexible or rigid) is often used but its efficacy and medium term impact is not studied or described. Hyperattenuated mucus plugging may be more difficult to relieve and is prone to relapse [58], and the most effective means of removal of hyperattenuated mucus and preventing relapse is not known.

Reduction of the fungal burden

Ketoconazole and inhaled natamycin were unsuccessful therapies for ABPA and have not been attempted for SAFS. There are no published data about nebulised amphotericin B for either ABPA or SAFS. Itraconazole was the first orally active agent against *Aspergillus* species and many other filamentous fungi. Two randomized placebo-controlled studies which enrolled a total of 84 patients both demonstrated benefit compared with placebo [201,202]. However the outcome parameters used in the studies were different and were composite in the first study. Pooled analysis showed that itraconazole reduced IgE levels by >25% when compared to placebo but did not significantly improve lung function. In the earlier, slightly larger study, the overall response rate was 60%, with a number needed to treat (NNT) of 3.58, consistent with a very effective intervention. Patients with ABPA may have continuing symptoms (asthma, bronchiectasis) as well as acute exacerbations. These were not separated out in terms of responses. Our own extensive experience is consistent with a range of responses, from dramatic through minor improvement to a complete lack of effect; the reasons for this response heterogeneity, despite adequate drug concentrations in blood, is

unclear. Relapse after conclusion of active therapy was not reported in either study. The frequency of acute exacerbations is not well known in ABPA, although was infrequent (16 of 41 (39%) over a mean period of 43 months) in a group of patients without bronchiectasis [193].

Fluconazole and itraconazole are suggested as possible effective treatments in the *Tricophyton* asthma [203], a phenotype of uncertain frequency. Fluconazole was effective in improving asthma in a small RCT in moderately severe asthmatics sensitized to *Trichophyton*, many with cutaneous fungal infection [204]. This has never been followed up. The use of terbinafine, which is active against *Trichophyton* spp., is only anecdotal [205]. The relative efficacy of fluconazole in those with multiple fungal sensitisation is not known, and is predicted to be inferior to itraconazole as it has a much more narrow spectrum of activity.

A broader approach was subsequently trialled using the wider spectrum agent itraconazole in SAFS patients sensitized to one or more of 7 fungi [27]. Compared to placebo, those treated with itraconazole over 8 months had a significant benefit in the primary endpoint of quality of life, as well as a fall in total IgE and marginal improvements in morning peak expiratory flow rate. The study was small (58 patients) and the confidence intervals just overlapped, so a replication study is warranted. Other data suggested a similar effect, but with some different parameters of response including reduced peripheral eosinophilia and improved FEV1 [206]. As with ABPA, our clinical experience suggestes some dramatic responses and some failures, with no clear cut reason discerbable for the variable response. While the effect size was similar to that seen in ABPA (NNT 3.22), the place of itraconazole in the management of severe asthma is not yet well defined.

The newer triazoles voriconazole and posaconazole have been effective in some patients with ABPA and SAFS, primarily in cystic fibrosis, but latterly in asthma [29]. Most experience to date has been in those intolerant to itraconazole or in itraconazole failures. Two remarkable findings in this retrospective review however were: (1) the large numbers of patients who were able to come off oral corticosteroids and substantially reduce their corticosteroid burden, and (2) that a third of the patients had their asthma severity downgraded from severe to moderate. These findings suggest a powerful impact in a proportion of ABPA and SAFS patients, which requires additional study (such as ATCF study, which is underway and aims to evaluate the rates of converting sputum cultures for *Aspergillus* from positive to negative, for itraconazole and voriconazole in a large prospective controlled clinical trial of CF patients). Unfortunately, the recently published EVITA3 study failed to show improvement in quality of life or the number of severe exacerbations during 9 months after termination of a 3 months' course of voriconazole in refractory asthma associated with sensitization to *A. fumigatus* [207]. Voriconazole 200 mg bd was given for only three months and while there was a significant reduction in the degree of *A. fumigatus* colonization in the voriconazole group the drug did not eradicate the mould from the sputum during the treatment period and rates of positive culture returned to baseline within a few months of stopping treatment. The placebo group had a surprisingly good improvement rate, and quality of life is often adversely affected by voriconazole, because of side effects.

As the bioavailability of itraconazole is variable and inadequate exposure is associated with higher clinical failure rates, therapeutic monitoring is required and is often not done or available. Variable absorption from different formulations of itraconazole contributes to the difficulties of maintaining consistent exposure and therefore effect. The penetration of

itraconazole, voriconazole and posaconazole into sputum and extracellular lung fluid (ELF) is variable and often low, despite adequate serum concentrations, for unclear reasons [208]. Voriconazole administered IV yielded high mean ELF concentrations (range 10.1-48.3 mg/L) and a high penetration ratio (7.1). In contrast, mean ELF concentrations of oral itraconazole and posaconazole were 0.2-1.9 mg/L, and the ELF penetration ratios were <1 [209]. However both itraconazole and posaconazole are concentrated inside alveolar cells.

The optimum usage of itraconazole and other oral azoles for therapy in ABPA is unclear presently, despite the 2 RCTs described above. Monotherapy of itraconazole in ABPA is possible but not studied. A randomized controlled trial comparing monotherapy of itraconazole versus prednisolone in ABPA (MIPA study; clinical trials.gov; NCT01321827) is underway, which aims to answer this question. The duration of therapy and frequency of relapse is unclear in ABPA, partly because exacerbation rates are infrequent. Reversion to pre-therapy status within 4 months was almost universal in SAFS patients who discontinued itraconazole therapy [27]. A recent study examining the impact of itraconazole on Vitamin D receptor expression in CF patients suggests a prolonged effect of down-regulation, lasting significantly longer than itraconazole therapy, possibly indicating that after successful therapy a sustained effect may be found in some patients [210].

The negative effects of itraconazole and possibly other azoles, other than their own direct toxicities, include a marked reduction in inhaled corticosteroid metabolism (with corresponding boosting of exposure) and azole resistance in *A.fumigatus* [211,212]. In a UK study, profound boosting of systemic corticosteroid exposure with inhaled corticosteroids was seen in 50% of patients treated with itraconazole, which reversed after 8 months [27]. Interactions between both budesonide and fluticasone and itraconazole are documented, and are likely with voriconazole and posaconazole, although may be less marked. Ciclosonide and beclometasone do not appear to interact with azoles. Adrenal failure is however described, typically after months or years of dual therapy.

Control of bacterial infection

A high frequency of respiratory bacterial infections is common in many of these patients. Most are presumed rather than documented, but bronchitis attributable to *Streptococcus pneumoniae* and *Haemophilus influenzae* is common, especially in those with bronchiectasis [213]. Long-term administration of azithromycin or doxycycline can alleviate some of these infections. Low specific antibodies to these bacteria are common in these patients and occasionally this is the presentation of hypogammaglobulinaemia. Immunisation with Pneumovax or Prevnar 13 to prevent *S. pneumoniae* infection and Menitorix to prevent *H. influenzae* is often done, but not well studied [214-217]. Individual immunisation responses are highly variable, and often poor to Pneumovax. Reduction of exacerbations and recurrent infection has a major benefit for patients, and reduces corticosteroid courses.

Treatment and general management research priorities

- *Additional studies on avoidance strategies to prevent fungal sensitisation in established asthma and course/exacerbations of fungi-related asthma and conditions related to incidental fungal exposures.*
- *The mechanism of benefit of antifungal therapy needs clarification, with development of biomarkers for response assessment.*

- *RCTs of voriconazole, posaconazole, isavuconazole and nebulized amphotericin B need to be conducted in ABPA and SAFS.*
- *Studies on antifungal and anti-IgE treatments in SAFS*
- *The role of adjunctive therapies such as macrolide, antibiotic prophylaxis, mucus reduction, the efficacy and impact of anti-*Pseudomonas *therapy needs to be assessed.*
- *The avoidance and management of azole/corticosteroid combinations needs addressing.*
- *The dual interaction of antifungal development of azole resistance and acquisition of a resistant strain on therapeutic impact requires addressing, given the increasing frequency of azole resistance in* A. fumigatus.
- *The optimal means and impact of reducing amplification of inflammation, through prevention of bacterial and viral intercurrent infection.*

Some patients become infected and others colonised by *Pseudomonas aeruginosa*, usually those with bronchiectasis [218]. Strategies to eradicate this with intravenous, dual antibiotic therapy are commonly used, without any clinical trial basis. The standard registered doses of ciprofloxacin are inadequate for most patients [219,220] and high doses such as 500-750 mg three times daily are more efficient. Newer approaches include oral high dose ciprofloxacin combined with nebulized colistin, following the lead of the cystic fibrosis community [221]. Data are lacking on impact and efficacy.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

All authors were involved in the Task Force discussions and contributed to writing the document. All authors read and approved the final manuscript.

Acknowledgment

References

1. Hardin BD, Kelman BJ, Saxon A: **Adverse human health effects associated with molds in the indoor environment.** *J Occup Environ Med* 2003, **45:**470–478.

2. Bush RK, Portnoy JM, Saxon A, Terr AI, Wood RA: **The medical effects of mold exposure.** *J Allergy Clin Immunol* 2006, **117:**326–333.

3. Stevens DA, Moss RB, Kurup VP, Knutsen AP, Greenberger P, Judson MA, *et al*: **Allergic bronchopulmonary aspergillosis in cystic fibrosis–state of the art: cystic fibrosis foundation consensus conference.** *Clin Infect Dis* 2003, **37**(Suppl 3)**:**S225–S264.

4. Knutsen AP, Bush RK, Demain JG, Denning DW, Dixit A, Fairs A, *et al*: **Fungi and allergic lower respiratory tract diseases.** *J Allergy Clin Immunol* 2012, **129:**280–291. quiz 292-283.

5. Agarwal R, Chakrabarti A, Shah A, Gupta D, Meis JF, Guleria R, *et al*: **Allergic bronchopulmonary aspergillosis: review of literature and proposal of new diagnostic and classification criteria.** *Clin Exp Allergy* 2013, **43:**850–873.

6. Pifer LL, Hughes WT, Stagno S, Woods D: **Pneumocystis carinii infection: evidence for high prevalence in normal and immunosuppressed children.** *Pediatrics* 1978, **61:**35–41.

7. Hedayati MT, Pasqualotto AC, Warn PA, Bowyer P, Denning DW: **Aspergillus flavus: human pathogen, allergen and mycotoxin producer.** *Microbiology* 2007, **153:**1677–1692.

8. Johansson SG, Hourihane JO, Bousquet J, Bruijnzeel-Koomen C, Dreborg S, Haahtela T, *et al*: **A revised nomenclature for allergy: an EAACI position statement from the EAACI nomenclature task force.** *Allergy* 2001, **56:**813–824.

9. Pirofski LA, Casadevall A: **The damage-response framework of microbial pathogenesis and infectious diseases.** *Adv Exp Med Biol* 2008, **635:**135–146.

10. Hargreave FE, Nair P: **The definition and diagnosis of asthma.** *Clin Exp Allergy* 2009, **39:**1652–1658.

11. Haldar P, Pavord ID, Shaw DE, Berry MA, Thomas M, Brightling CE, *et al*: **Cluster analysis and clinical asthma phenotypes.** *Am J Respir Crit Care Med* 2008, **178:**218–224.

12. Lotvall J, Akdis CA, Bacharier LB, Bjermer L, Casale TB, Custovic A, *et al*: **Asthma endotypes: a new approach to classification of disease entities within the asthma syndrome.** *J Allergy Clin Immunol* 2011, **127:**355–360.

13. Hinson KF, Moon AJ, Plummer NS: **Broncho-pulmonary aspergillosis; a review and a report of eight new cases.** *Thorax* 1952, **7:**317–333.

14. Greenberger PA, Patterson R: **Allergic bronchopulmonary aspergillosis and the evaluation of the patient with asthma.** *J Allergy Clin Immunol* 1988, **81:**646–650.

15. McCarthy DS, Pepys J: **Allergic broncho-pulmonary aspergillosis: clinical immunology. 2: skin, nasal and bronchial tests.** *Clin Allergy* 1971, **1:**415–432.

16. Rosenberg M, Patterson R, Mintzer R, Cooper BJ, Roberts M, Harris KE: **Clinical and immunologic criteria for the diagnosis of allergic bronchopulmonary aspergillosis.** *Ann Intern Med* 1977, **86:**405–414.

17. McCarthy DS, Pepys J: **Allergic broncho-pulmonary aspergillosis.** *Clin Allergy* 1971, **1:**261–281.

18. Baxter CG, Dunn G, Jones AM, Webb K, Gore R, Richardson MD, *et al*: **Novel immunologic classification of aspergillosis in adult cystic fibrosis.** *J Allergy Clin Immunol* 2013, **132:**560–566. e510.

19. Agarwal R, Maskey D, Aggarwal AN, Saikia B, Garg M, Gupta D, *et al*: **Diagnostic performance of various tests and criteria employed in allergic bronchopulmonary aspergillosis: a latent class analysis.** *PLoS One* 2013, **8:**e61105.

20. Agarwal R, Khan A, Aggarwal AN, Varma N, Garg M, Saikia B, *et al*: **Clinical relevance of peripheral blood eosinophil count in allergic bronchopulmonary aspergillosis.** *J Infect Public Health* 2011, **4:**235–243.

21. Woellner C, Gertz EM, Schaffer AA, Lagos M, Perro M, Glocker EO, *et al*: **Mutations in STAT3 and diagnostic guidelines for hyper-IgE syndrome.** *J Allergy Clin Immunol* 2010, **125:**424–432. e428.

22. Chrdle A, Mustakim S, Bright-Thomas RJ, Baxter CG, Felton T, Denning DW: **Aspergillus bronchitis without significant immunocompromise.** *Ann N Y Acad Sci* 2012, **1272:**73–85.

23. Denning DW, Riniotis K, Dobrashian R, Sambatakou H: **Chronic cavitary and fibrosing pulmonary and pleural aspergillosis: case series, proposed nomenclature change, and review.** *Clin Infect Dis* 2003, **37**(Suppl 3)**:**S265–S280.

24. Fairs A, Agbetile J, Hargadon B, Bourne M, Monteiro WR, Brightling CE, *et al*: **IgE sensitization to Aspergillus fumigatus is associated with reduced lung function in asthma.** *Am J Respir Crit Care Med* 2010, **182:**1362–1368.

25. Chowdhary A, Agarwal K, Kathuria S, Gaur SN, Randhawa HS, Meis JF: **Allergic bronchopulmonary mycosis due to fungi other than Aspergillus: a global overview.** *Crit Rev Microbiol* 2014, **40:**30–48.

26. Denning DW, O'Driscoll BR, Hogaboam CM, Bowyer P, Niven RM: **The link between fungi and severe asthma: a summary of the evidence.** *Eur Respir J* 2006, **27:**615–626.

27. Denning DW, O'Driscoll BR, Powell G, Chew F, Atherton GT, Vyas A, *et al*: **Randomized controlled trial of oral antifungal treatment for severe asthma with fungal sensitization: The Fungal Asthma Sensitization Trial (FAST) study.** *Am J Respir Crit Care Med* 2009, **179:**11–18.

28. Bush A, Zar HJ: **WHO universal definition of severe asthma.** *Curr Opin Allergy Clin Immunol* 2011, **11:**115–121.

29. Chishimba L, Niven RM, Cooley J, Denning DW: **Voriconazole and posaconazole improve asthma severity in allergic bronchopulmonary aspergillosis and severe asthma with fungal sensitization.** *J Asthma* 2012, **49:**423–433.

30. Carvalho A, Pasqualotto AC, Pitzurra L, Romani L, Denning DW, Rodrigues F: **Polymorphisms in toll-like receptor genes and susceptibility to pulmonary aspergillosis.** *J Infect Dis* 2008, **197:**618–621.

31. Schubert MS: **Allergic fungal sinusitis: pathophysiology, diagnosis and management.** *Med Mycol* 2009, **47**(Suppl 1)**:**S324–S330.

32. Chakrabarti A, Denning DW, Ferguson BJ, Ponikau J, Buzina W, Kita H, *et al*: **Fungal rhinosinusitis: a categorization and definitional schema addressing current controversies.** *Laryngoscope* 2009, **119:**1809–1818.

33. Bafadhel M, McKenna S, Agbetile J, Fairs A, Desai D, Mistry V, *et al*: **Aspergillus.** *Eur Respir J* 2013, **43:**64–71.

34. Menzies D, Holmes L, McCumesky G, Prys-Picard C, Niven R: **Aspergillus sensitization is associated with airflow limitation and bronchiectasis in severe asthma.** *Allergy* 2011, **66:**679–685.

35. Stern DA, Morgan WJ, Halonen M, Wright AL, Martinez FD: **Wheezing and bronchial hyper-responsiveness in early childhood as predictors of newly diagnosed asthma in early adulthood: a longitudinal birth-cohort study.** *Lancet* 2008, **372:**1058–1064.

36. Green BJ, Mitakakis TZ, Tovey ER: **Allergen detection from 11 fungal species before and after germination.** *J Allergy Clin Immunol* 2003, **111:**285–289.

37. Aimanianda V, Bayry J, Bozza S, Kniemeyer O, Perruccio K, Elluru SR, *et al*: **Surface hydrophobin prevents immune recognition of airborne fungal spores.** *Nature* 2009, **460:**1117–1121.

38. Green BJ, Sercombe JK, Tovey ER: **Fungal fragments and undocumented conidia function as new aeroallergen sources.** *J Allergy Clin Immunol* 2005, **115:**1043–1048.

39. Ward GW Jr, Karlsson G, Rose G, Platts-Mills TA: **Trichophyton asthma: sensitisation of bronchi and upper airways to dermatophyte antigen.** *Lancet* 1989, **1:**859–862.

40. Matsuoka H, Niimi A, Matsumoto H, Ueda T, Takemura M, Yamaguchi M, *et al*: **Specific IgE response to trichophyton and asthma severity.** *Chest* 2009, **135:**898–903.

41. Escalante MT, Sanchez-Borges M, Capriles-Hulett A, Belfort E, Di Biagio E, Gonzalez-Aveledo L: **Trichophyton-specific IgE in patients with dermatophytosis is not associated with aeroallergen sensitivity.** *J Allergy Clin Immunol* 2000, **105:**547–551.

42. Mungan D, Bavbek S, Peksari V, Celik G, Gugey E, Misirligil Z: **Trichophyton sensitivity in allergic and nonallergic asthma.** *Allergy* 2001, **56:**558–562.

43. O'Driscoll BR, Powell G, Chew F, Niven RM, Miles JF, Vyas A, *et al*: **Comparison of skin prick tests with specific serum immunoglobulin E in the diagnosis of fungal sensitization in patients with severe asthma.** *Clin Exp Allergy* 2009, **39:**1677–1683.

44. Imbeau SA, Cohen M, Reed CE: **Allergic bronchopulmonary aspergillosis in infants.** *Am J Dis Child* 1977, **131:**1127–1130.

45. Wang JL, Patterson R, Mintzer R, Roberts M, Rosenberg M: **Allergic bronchopulmonary aspergillosis in pediatric practice.** *J Pediatr* 1979, **94:**376–381.

46. Chetty A, Bhargava S, Jain RK: **Allergic bronchopulmonary aspergillosis in Indian children with bronchial asthma.** *Ann Allergy* 1985, **54:**46–49.

47. Patterson R, Greenberger PA, Halwig JM, Liotta JL, Roberts M: **Allergic bronchopulmonary aspergillosis: natural history and classification of early disease by serologic and roentgenographic studies.** *Arch Intern Med* 1986, **146:**916–918.

48. Patterson R, Greenberger PA, Radin RC, Roberts M: **Allergic bronchopulmonary aspergillosis: staging as an aid to management.** *Ann Intern Med* 1982, **96:**286–291.

49. Greene R: **The pulmonary aspergilloses: three distinct entities or a spectrum of disease.** *Radiology* 1981, **140:**527–530.

50. Chetty A: **Pathology of allergic bronchopulmonary aspergillosis.** *Front Biosci* 2003, **8:**e110–e114.

51. Smith NL, Denning DW: **Underlying conditions in chronic pulmonary aspergillosis including simple aspergilloma.** *Eur Respir J* 2011, **37:**865–872.

52. Patterson R, Greenberger PA, Harris KE: **Allergic bronchopulmonary aspergillosis.** *Chest* 2000, **118:**7–8.

53. Kumar R: **Mild, moderate, and severe forms of allergic bronchopulmonary aspergillosis: a clinical and serologic evaluation.** *Chest* 2003, **124:**890–892.

54. Greenberger PA, Miller TP, Roberts M, Smith LL: **Allergic bronchopulmonary aspergillosis in patients with and without evidence of bronchiectasis.** *Ann Allergy* 1993, **70:**333–338.

55. Mitchell TA, Hamilos DL, Lynch DA, Newell JD: **Distribution and severity of bronchiectasis in allergic bronchopulmonary aspergillosis (ABPA).** *J Asthma* 2000, **37:**65–72.

56. Agarwal R, Garg M, Aggarwal AN, Saikia B, Gupta D, Chakrabarti A: **Serologic allergic bronchopulmonary aspergillosis (ABPA-S): long-term outcomes.** *Respir Med* 2012, **106:**942–947.

57. Agarwal R, Gupta D, Aggarwal AN, Behera D, Jindal SK: **Allergic bronchopulmonary aspergillosis: lessons from 126 patients attending a chest clinic in north India.** *Chest* 2006, **130:**442–448.

58. Agarwal R, Gupta D, Aggarwal AN, Saxena AK, Chakrabarti A, Jindal SK: **Clinical significance of hyperattenuating mucoid impaction in allergic bronchopulmonary aspergillosis: an analysis of 155 patients.** *Chest* 2007, **132:**1183–1190.

59. Gefter WB, Epstein DM, Miller TW: **Allergic bronchopulmonary aspergillosis: less common patterns.** *Radiology* 1981, **140:**307–312.

60. Israel RH, Poe RH, Bomba PA, Gross RA: **The rapid development of an aspergilloma secondary to allergic bronchopulmonary aspergillosis.** *Am J Med Sci* 1980, **280:**41–44.

61. Riley DJ, Mackenzie JW, Uhlman WE, Edelman NH: **Allergic bronchopulmonary aspergillosis: evidence of limited tissue invasion.** *Am Rev Respir Dis* 1975, **111:**232–236.

62. Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, Aboyans V, *et al*: **Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the global burden of disease study 2010.** *Lancet* 2012, **380:**2095–2128.

63. Vos T, Flaxman AD, Naghavi M, Lozano R, Michaud C, Ezzati M, *et al*: **Years lived with disability (YLDs) for 1160 sequelae of 289 diseases and injuries 1990-2010: a systematic analysis for the global burden of disease study 2010.** *Lancet* 2012, **380:**2163–2196.

64. Murray CJ, Vos T, Lozano R, Naghavi M, Flaxman AD, Michaud C, *et al*: **Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: a systematic analysis for the global burden of disease study 2010.** *Lancet* 2012, **380:**2197–2223.

65. Denning DW, Pleuvry A, Cole DC: **Global burden of allergic bronchopulmonary aspergillosis with asthma and its complication chronic pulmonary aspergillosis in adults.** *Med Mycol* 2013, **51:**361–370.

66. Hulin M, Moularat S, Kirchner S, Robine E, Mandin C, Annesi-Maesano I: **Positive associations between respiratory outcomes and fungal index in rural inhabitants of a representative sample of French dwellings.** *Int J Hyg Environ Health* 2013, **216:**155–162.

67. Reponen T, Lockey J, Bernstein DI, Vesper SJ, Levin L, Khurana Hershey GK, *et al*: **Infant origins of childhood asthma associated with specific molds.** *J Allergy Clin Immunol* 2012, **130:**639–644. e635.

68. Reponen T, Vesper S, Levin L, Johansson E, Ryan P, Burkle J, *et al*: **High environmental relative moldiness index during infancy as a predictor of asthma at 7 years of age.** *Ann Allergy Asthma Immunol* 2011, **107:**120–126.

69. Karjalainen A, Kurppa K, Martikainen R, Klaukka T, Karjalainen J: **Work is related to a substantial portion of adult-onset asthma incidence in the Finnish population.** *Am J Respir Crit Care Med* 2001, **164:**565–568.

70. Park JH, Cox-Ganser JM, Kreiss K, White SK, Rao CY: **Hydrophilic fungi and ergosterol associated with respiratory illness in a water-damaged building.** *Environ Health Perspect* 2008, **116:**45–50.

71. Piipari R, Keskinen H: **Agents causing occupational asthma in Finland in 1986-2002: cow epithelium bypassed by moulds from moisture-damaged buildings.** *Clin Exp Allergy* 2005, **35:**1632–1637.

72. McDonald JC, Keynes HL, Meredith SK: **Reported incidence of occupational asthma in the United Kingdom, 1989-97.** *Occup Environ Med* 2000, **57:**823–829.

73. Quirce S, Cuevas M, Diez-Gomez M, Fernandez-Rivas M, Hinojosa M, Gonzalez R, *et al*: **Respiratory allergy to Aspergillus-derived enzymes in bakers' asthma.** *J Allergy Clin Immunol* 1992, **90:**970–978.

74. Moreno-Ancillo A, Dominguez-Noche C, Gil-Adrados AC, Cosmes PM: **Bread eating induced oral angioedema due to alpha-amylase allergy.** *J Investig Allergol Clin Immunol* 2004, **14:**346–347.

75. Yamamoto N, Bibby K, Qian J, Hospodsky D, Rismani-Yazdi H, Nazaroff WW, *et al*: **Particle-size distributions and seasonal diversity of allergenic and pathogenic fungi in outdoor air.** *Isme J* 2012, **6:**1801–1811.

76. Atkinson RW, Strachan DP, Anderson HR, Hajat S, Emberlin J: **Temporal associations between daily counts of fungal spores and asthma exacerbations.** *Occup Environ Med* 2006, **63:**580–590.

77. Newson R, Strachan D, Corden J, Millington W: **Fungal and other spore counts as predictors of admissions for asthma in the Trent region.** *Occup Environ Med* 2000, **57:**786–792.

78. Black PN, Udy AA, Brodie SM: **Sensitivity to fungal allergens is a risk factor for life-threatening asthma.** *Allergy* 2000, **55:**501–504.

79. O'Hollaren MT, Yunginger JW, Offord KP, Somers MJ, O'Connell EJ, Ballard DJ, *et al*: **Exposure to an aeroallergen as a possible precipitating factor in respiratory arrest in young patients with asthma.** *N Engl J Med* 1991, **324:**359–363.

80. Delfino RJ, Zeiger RS, Seltzer JM, Street DH, Matteucci RM, Anderson PR, *et al*: **The effect of outdoor fungal spore concentrations on daily asthma severity.** *Environ Health Perspect* 1997, **105:**622–635.

81. Pulimood TB, Corden JM, Bryden C, Sharples L, Nasser SM: **Epidemic asthma and the role of the fungal mold Alternaria alternata.** *J Allergy Clin Immunol* 2007, **120:**610–617.

82. Wong GW, Chow CM: **Childhood asthma epidemiology: insights from comparative studies of rural and urban populations.** *Pediatr Pulmonol* 2008, **43:**107–116.

83. Agarwal RDD, Gupta D, Chakrabarti A: *A questionnaire-based study on the role of environmental factors in allergic bronchopulmonary aspergillosis.* India: Lung; 2014. in press.

84. Flamant-Hulin M, Annesi-Maesano I, Caillaud D: **Relationships between molds and asthma suggesting non-allergic mechanisms: a rural–urban comparison.** *Pediatr Allergy Immunol* 2013, **24:**345–351.

85. Ren P, Jankun TM, Belanger K, Bracken MB, Leaderer BP: **The relation between fungal propagules in indoor air and home characteristics.** *Allergy* 2001, **56:**419–424.

86. Zock JP, Jarvis D, Luczynska C, Sunyer J, Burney P: **Housing characteristics, reported mold exposure, and asthma in the European community respiratory health survey.** *J Allergy Clin Immunol* 2002, **110:**285–292.

87. Norback D, Bjornsson E, Janson C, Palmgren U, Boman G: **Current asthma and biochemical signs of inflammation in relation to building dampness in dwellings.** *Int J Tuberc Lung Dis* 1999, **3:**368–376.

88. Agbetile J, Fairs A, Desai D, Hargadon B, Bourne M, Mutalithas K, *et al*: **Isolation of filamentous fungi from sputum in asthma is associated with reduced post-bronchodilator FEV1.** *Clin Exp Allergy* 2012, **42:**782–791.

89. Gergen PJ, Turkeltaub PC: **The association of individual allergen reactivity with respiratory disease in a national sample: data from the second National Health and Nutrition Examination Survey, 1976-80 (NHANES II).** *J Allergy Clin Immunol* 1992, **90:**579–588.

90. Salo PM, Arbes SJ Jr, Sever M, Jaramillo R, Cohn RD, London SJ, *et al*: **Exposure to Alternaria alternata in US homes is associated with asthma symptoms.** *J Allergy Clin Immunol* 2006, **118:**892–898.

91. Williamson IJ, Martin CJ, McGill G, Monie RD, Fennerty AG: **Damp housing and asthma: a case–control study.** *Thorax* 1997, **52:**229–234.

92. Wood RA, Eggleston PA, Lind P, Ingemann L, Schwartz B, Graveson S, *et al*: **Antigenic analysis of household dust samples.** *Am Rev Respir Dis* 1988, **137:**358–363.

93. Norback D, Bjornsson E, Janson C, Widstrom J, Boman G: **Asthmatic symptoms and volatile organic compounds, formaldehyde, and carbon dioxide in dwellings.** *Occup Environ Med* 1995, **52:**388–395.

94. Platt SD, Martin CJ, Hunt SM, Lewis CW: **Damp housing, mould growth, and symptomatic health state.** *BMJ* 1989, **298:**1673–1678.

95. Jaakkola MS, Ieromnimon A, Jaakkola JJ: **Are atopy and specific IgE to mites and molds important for adult asthma?** *J Allergy Clin Immunol* 2006, **117:**642–648.

96. Fairs A, Agbetile J, Bourne M, Hargadon B, Monteiro WR, Morley JP, *et al*: **Isolation of Aspergillus fumigatus from sputum is associated with elevated airborne levels in homes of patients with asthma.** *Indoor Air* 2013, **23:**275–284.

97. Mannes GP, van der Heide S, van Aalderen WM, Gerritsen J: **Itraconazole and allergic bronchopulmonary aspergillosis in twin brothers with cystic fibrosis.** *Lancet* 1993, **341:**492.

98. Shah A, Kala J, Sahay S, Panjabi C: **Frequency of familial occurrence in 164 patients with allergic bronchopulmonary aspergillosis.** *Ann Allergy Asthma Immunol* 2008, **101:**363–369.

99. Bains SN, Judson MA: **Allergic bronchopulmonary aspergillosis.** *Clin Chest Med* 2012, **33:**265–281.

100. Armougom F, Bittar F, Stremler N, Rolain JM, Robert C, Dubus JC, *et al*: **Microbial diversity in the sputum of a cystic fibrosis patient studied with 16S rDNA pyrosequencing.** *Eur J Clin Microbiol Infect Dis* 2009, **28:**1151–1154.

101. Bittar F, Richet H, Dubus JC, Reynaud-Gaubert M, Stremler N, Sarles J, *et al*: **Molecular detection of multiple emerging pathogens in sputa from cystic fibrosis patients.** *PLoS One* 2008, **3:**e2908.

102. Chabe M, Aliouat-Denis CM, Delhaes L, el Aliouat M, Viscogliosi E, Dei-Cas E: **Pneumocystis: from a doubtful unique entity to a group of highly diversified fungal species.** *FEMS Yeast Res* 2011, **11:**2–17.

103. Cottier F, Pavelka N: **Complexity and dynamics of host-fungal interactions.** *Immunol Res* 2012, **53:**127–135.

104. Delhaes L, Monchy S, Frealle E, Hubans C, Salleron J, Leroy S, *et al*: **The airway microbiota in cystic fibrosis: a complex fungal and bacterial community–implications for therapeutic management.** *PLoS One* 2012, **7:**e36313.

105. Santamaria M, Fosso B, Consiglio A, De Caro G, Grillo G, Licciulli F, *et al*: **Reference databases for taxonomic assignment in metagenomics.** *Brief Bioinform* 2012, **13:**682–695.

106. Charlson ES, Diamond JM, Bittinger K, Fitzgerald AS, Yadav A, Haas AR, *et al*: **Lung-enriched organisms and aberrant bacterial and fungal respiratory microbiota after lung transplant.** *Am J Respir Crit Care Med* 2012, **186:**536–545.

107. Hilty M, Burke C, Pedro H, Cardenas P, Bush A, Bossley C, *et al*: **Disordered microbial communities in asthmatic airways.** *PLoS One* 2010, **5:**e8578.

108. van Woerden HC, Gregory C, Brown R, Marchesi JR, Hoogendoorn B, Matthews IP: **Differences in fungi present in induced sputum samples from asthma patients and non-atopic controls: a community based case control study.** *BMC Infect Dis* 2013, **13:**69.

109. Fittipaldi M, Nocker A, Codony F: **Progress in understanding preferential detection of live cells using viability dyes in combination with DNA amplification.** *J Microbiol Methods* 2012, **91:**276–289.

110. Rogers GB, Cuthbertson L, Hoffman LR, Wing PA, Pope C, Hooftman DA, *et al*: **Reducing bias in bacterial community analysis of lower respiratory infections.** *Isme J* 2013, **7:**697–706.

111. Pashley CH, Fairs A, Free RC, Wardlaw AJ: **DNA analysis of outdoor air reveals a high degree of fungal diversity, temporal variability, and genera not seen by spore morphology.** *Fungal Biol* 2012, **116:**214–224.

112. Vautier S, MacCallum DM, Brown GD: **C-type lectin receptors and cytokines in fungal immunity.** *Cytokine* 2012, **58:**89–99.

113. Romani L, Puccetti P: **Immune regulation and tolerance to fungi in the lungs and skin.** *Chem Immunol Allergy* 2008, **94:**124–137.

114. Hogan C, Denning DW: **Allergic bronchopulmonary aspergillosis and related allergic syndromes.** *Semin Respir Crit Care Med* 2011, **32:**682–692.

115. Roilides E, Simitsopoulou M: **Local innate host response and filamentous fungi in patients with cystic fibrosis.** *Med Mycol* 2010, **48**(Suppl 1)**:**S22–S31.

116. Chotirmall SH, Greene CM, McElvaney NG: **Candida species in cystic fibrosis: a road less travelled.** *Med Mycol* 2010, **48**(Suppl 1)**:**S114–S124.

117. Hogaboam CM, Carpenter KJ, Schuh JM, Buckland KF: **Aspergillus and asthma–any link?** *Med Mycol* 2005, **43**(Suppl 1)**:**S197–S202.

118. Brown GD: **Innate antifungal immunity: the key role of phagocytes.** *Annu Rev Immunol* 2011, **29:**1–21.

119. Urban CF, Ermert D, Schmid M, Abu-Abed U, Goosmann C, Nacken W, *et al*: **Neutrophil extracellular traps contain calprotectin, a cytosolic protein complex involved in host defense against Candida albicans.** *PLoS Pathog* 2009, **5:**e1000639.

120. Bouzani M, Ok M, McCormick A, Ebel F, Kurzai O, Morton CO, *et al*: **Human NK cells display important antifungal activity against Aspergillus fumigatus, which is directly mediated by IFN-gamma release.** *J Immunol* 2011, **187:**1369–1376.

121. Seider K, Brunke S, Schild L, Jablonowski N, Wilson D, Majer O, *et al*: **The facultative intracellular pathogen Candida glabrata subverts macrophage cytokine production and phagolysosome maturation.** *J Immunol* 2011, **187:**3072–3086.

122. McKenzie CG, Koser U, Lewis LE, Bain JM, Mora-Montes HM, Barker RN, *et al*: **Contribution of Candida albicans cell wall components to recognition by and escape from murine macrophages.** *Infect Immun* 2010, **78:**1650–1658.

123. Hasenberg M, Behnsen J, Krappmann S, Brakhage A, Gunzer M: **Phagocyte responses towards Aspergillus fumigatus.** *Int J Med Microbiol* 2011, **301:**436–444.

124. Balloy V, Chignard M: **The innate immune response to Aspergillus fumigatus.** *Microbes Infect* 2009, **11:**919–927.

125. Hartl D, Buckland KF, Hogaboam CM: **Chemokines in allergic aspergillosis–from animal models to human lung diseases.** *Inflamm Allergy Drug Targets* 2006, **5:**219–228.

126. Hartl D: **Immunological mechanisms behind the cystic fibrosis-ABPA link.** *Med Mycol* 2009, **47**(Suppl 1)**:**S183–S191.

127. Hartl D, Latzin P, Zissel G, Krane M, Krauss-Etschmann S, Griese M: **Chemokines indicate allergic bronchopulmonary aspergillosis in patients with cystic fibrosis.** *Am J Respir Crit Care Med* 2006, **173:**1370–1376.

128. Latzin P, Hartl D, Regamey N, Frey U, Schoeni MH, Casaulta C: **Comparison of serum markers for allergic bronchopulmonary aspergillosis in cystic fibrosis.** *Eur Respir J* 2008, **31:**36–42.

129. McAleer JP, Kolls JK: **Mechanisms controlling Th17 cytokine expression and host defense.** *J Leukoc Biol* 2011, **90:**263–270.

130. Glocker EO, Grimbacher B: **Mucosal antifungal defence: IL-17 signalling takes centre stage.** *Immunol Cell Biol* 2011, **89:**823–825.

131. Puel A, Cypowyj S, Bustamante J, Wright JF, Liu L, Lim HK, *et al*: **Chronic mucocutaneous candidiasis in humans with inborn errors of interleukin-17 immunity.** *Science* 2011, **332:**65–68.

132. Zelante T, Iannitti RG, De Luca A, Arroyo J, Blanco N, Servillo G, *et al*: **Sensing of mammalian IL-17A regulates fungal adaptation and virulence.** *Nat Commun* 2012, **3:**683.

133. Zelante T, Bozza S, De Luca A, D'Angelo C, Bonifazi P, Moretti S, *et al*: **Th17 cells in the setting of Aspergillus infection and pathology.** *Med Mycol* 2009, **47**(Suppl 1)**:**S162–S169.

134. Romani L, Fallarino F, De Luca A, Montagnoli C, D'Angelo C, Zelante T, *et al*: **Defective tryptophan catabolism underlies inflammation in mouse chronic granulomatous disease.** *Nature* 2008, **451:**211–215.

135. Romani L, Zelante T, De Luca A, Fallarino F, Puccetti P: **IL-17 and therapeutic kynurenines in pathogenic inflammation to fungi.** *J Immunol* 2008, **180:**5157–5162.

136. Romani L: **Immunity to fungal infections.** *Nat Rev Immunol* 2011, **11:**275–288.

137. De Ravin SS, Zarember KA, Long-Priel D, Chan KC, Fox SD, Gallin JI, *et al*: **Tryptophan/kynurenine metabolism in human leukocytes is independent of superoxide and is fully maintained in chronic granulomatous disease.** *Blood* 2010, **116:**1755–1760.

138. Jurgens B, Fuchs D, Reichenbach J, Heitger A: **Intact indoleamine 2,3-dioxygenase activity in human chronic granulomatous disease.** *Clin Immunol* 2010, **137:**1–4.

139. Kuijpers T, Lutter R: **Inflammation and repeated infections in CGD: two sides of a coin.** *Cell Mol Life Sci* 2012, **69:**7–15.

140. Rieber N, Hector A, Kuijpers T, Roos D, Hartl D: **Current concepts of hyperinflammation in chronic granulomatous disease.** *Clin Dev Immunol* 2012, **2012:**252460.

141. Hardison SE, Brown GD: **C-type lectin receptors orchestrate antifungal immunity.** *Nat Immunol* 2012, **13:**817–822.

142. Goodridge HS, Reyes CN, Becker CA, Katsumoto TR, Ma J, Wolf AJ, *et al*: **Activation of the innate immune receptor Dectin-1 upon formation of a 'phagocytic synapse'.** *Nature* 2011, **472:**471–475.

143. Faro-Trindade I, Willment JA, Kerrigan AM, Redelinghuys P, Hadebe S, Reid DM, *et al*: **Characterisation of innate fungal recognition in the lung.** *PLoS One* 2012, **7:**e35675.

144. Ferwerda B, Ferwerda G, Plantinga TS, Willment JA, van Spriel AB, Venselaar H, *et al*: **Human dectin-1 deficiency and mucocutaneous fungal infections.** *N Engl J Med* 2009, **361:**1760–1767.

145. Glocker EO, Hennigs A, Nabavi M, Schaffer AA, Woellner C, Salzer U, *et al*: **A homozygous CARD9 mutation in a family with susceptibility to fungal infections.** *N Engl J Med* 2009, **361:**1727–1735.

146. van de Veerdonk FL, Plantinga TS, Hoischen A, Smeekens SP, Joosten LA, Gilissen C, *et al*: **STAT1 mutations in autosomal dominant chronic mucocutaneous candidiasis.** *N Engl J Med* 2011, **365:**54–61.

147. Netea MG, Ferwerda G, van der Graaf CA, Van der Meer JW, Kullberg BJ: **Recognition of fungal pathogens by toll-like receptors.** *Curr Pharm Des* 2006, **12:**4195–4201.

148. Moreira AP, Cavassani KA, Ismailoglu UB, Hullinger R, Dunleavy MP, Knight DA, *et al*: **The protective role of TLR6 in a mouse model of asthma is mediated by IL-23 and IL-17A.** *J Clin Invest* 2011, **121:**4420–4432.

149. Cunha C, Romani L, Carvalho A: **Cracking the Toll-like receptor code in fungal infections.** *Expert Rev Anti Infect Ther* 2010, **8:**1121–1137.

150. Bellocchio S, Moretti S, Perruccio K, Fallarino F, Bozza S, Montagnoli C, *et al*: **TLRs govern neutrophil activity in aspergillosis.** *J Immunol* 2004, **173:**7406–7415.

151. Bochud PY, Chien JW, Marr KA, Leisenring WM, Upton A, Janer M, *et al*: **Toll-like receptor 4 polymorphisms and aspergillosis in stem-cell transplantation.** *N Engl J Med* 2008, **359:**1766–1777.

152. Kesh S, Mensah NY, Peterlongo P, Jaffe D, Hsu K, Van den Brink M, *et al*: **TLR1 and TLR6 polymorphisms are associated with susceptibility to invasive aspergillosis after allogeneic stem cell transplantation.** *Ann N Y Acad Sci* 2005, **1062:**95–103.

153. Knutsen AP, Kariuki B, Consolino JD, Warrier MR: **IL-4 alpha chain receptor (IL-4Ralpha) polymorphisms in allergic bronchopulmonary sspergillosis.** *Clin Mol Allergy* 2006, **4:**3.

154. Muller U, Piehler D, Stenzel W, Kohler G, Frey O, Held J, *et al*: **Lack of IL-4 receptor expression on T helper cells reduces T helper 2 cell polyfunctionality and confers resistance in allergic bronchopulmonary mycosis.** *Mucosal Immunol* 2012, **5:**299–310.

155. Brouard J, Knauer N, Boelle PY, Corvol H, Henrion-Caude A, Flamant C, *et al*: **Influence of interleukin-10 on Aspergillus fumigatus infection in patients with cystic fibrosis.** *J Infect Dis* 2005, **191:**1988–1991.

156. Vaid M, Kaur S, Sambatakou H, Madan T, Denning DW, Sarma PU: **Distinct alleles of mannose-binding lectin (MBL) and surfactant proteins A (SP-A) in patients with chronic cavitary pulmonary aspergillosis and allergic bronchopulmonary aspergillosis.** *Clin Chem Lab Med* 2007, **45:**183–186.

157. Miller PW, Hamosh A, Macek M Jr, Greenberger PA, MacLean J, Walden SM, *et al*: **Cystic fibrosis transmembrane conductance regulator (CFTR) gene mutations in allergic bronchopulmonary aspergillosis.** *Am J Hum Genet* 1996, **59:**45–51.

158. Marchand E, Verellen-Dumoulin C, Mairesse M, Delaunois L, Brancaleone P, Rahier JF, *et al*: **Frequency of cystic fibrosis transmembrane conductance regulator gene mutations and 5T allele in patients with allergic bronchopulmonary aspergillosis.** *Chest* 2001, **119:**762–767.

159. Agarwal R, Khan A, Aggarwal AN, Gupta D: **Link between CFTR mutations and ABPA: a systematic review and meta-analysis.** *Mycoses* 2012, **55:**357–365.

160. Aron Y, Bienvenu T, Hubert D, Dusser D, Dall'Ava J, Polla BS: **HLA-DR polymorphism in allergic bronchopulmonary aspergillosis.** *J Allergy Clin Immunol* 1999, **104:**891–892.

161. Knutsen AP, Slavin RG: **Allergic bronchopulmonary aspergillosis in asthma and cystic fibrosis.** *Clin Dev Immunol* 2011, **2011:**843763.

162. Simon-Nobbe B, Denk U, Poll V, Rid R, Breitenbach M: **The spectrum of fungal allergy.** *Int Arch Allergy Immunol* 2008, **145:**58–86.

163. Agarwal R, Aggarwal AN, Gupta D, Jindal SK: **Aspergillus hypersensitivity and allergic bronchopulmonary aspergillosis in patients with bronchial asthma: systematic review and meta-analysis.** *Int J Tuberc Lung Dis* 2009, **13:**936–944.

164. Crameri R, Hemmann S, Ismail C, Menz G, Blaser K: **Disease-specific recombinant allergens for the diagnosis of allergic bronchopulmonary aspergillosis.** *Int Immunol* 1998, **10:**1211–1216.

165. Hemmann S, Nikolaizik WH, Schoni MH, Blaser K, Crameri R: **Differential IgE recognition of recombinant Aspergillus fumigatus allergens by cystic fibrosis patients with allergic bronchopulmonary aspergillosis or Aspergillus allergy.** *Eur J Immunol* 1998, **28:**1155–1160.

166. Kurup VP, Banerjee B, Hemmann S, Greenberger PA, Blaser K, Crameri R: **Selected recombinant Aspergillus fumigatus allergens bind specifically to IgE in ABPA.** *Clin Exp Allergy* 2000, **30:**988–993.

167. Greenberger PA: **Allergic bronchopulmonary aspergillosis.** *J Allergy Clin Immunol* 2002, **110:**685–692.

168. Tillie-Leblond I, Tonnel AB: **Allergic bronchopulmonary aspergillosis.** *Allergy* 2005, **60:**1004–1013.

169. Agarwal R: **Allergic bronchopulmonary aspergillosis.** *Chest* 2009, **135:**805–826.

170. Pashley CH, Fairs A, Morley JP, Tailor S, Agbetile J, Bafadhel M, *et al*: **Routine processing procedures for isolating filamentous fungi from respiratory sputum samples may underestimate fungal prevalence.** *Med Mycol* 2012, **50:**433–438.

171. Borman AM, Palmer MD, Delhaes L, Carrere J, Favennec L, Ranque S, *et al*: **Lack of standardization in the procedures for mycological examination of sputum samples from CF patients: a possible cause for variations in the prevalence of filamentous fungi.** *Med Mycol* 2010, **48**(Suppl 1)**:**S88–S97.

172. Baxter CG, Jones AM, Webb K, Denning DW: **Homogenisation of cystic fibrosis sputum by sonication–an essential step for Aspergillus PCR.** *J Microbiol Methods* 2011, **85:**75–81.

173. Fraczek MG, Kirwan MB, Moore CB, Morris J, Denning DW, Richardson MD: **Volume dependency for culture of fungi from respiratory secretions and increased sensitivity of Aspergillus quantitative PCR.** *Mycoses* 2013, **57:**69–78.

174. Denning DW, Park S, Lass-Florl C, Fraczek MG, Kirwan M, Gore R, *et al*: **High-frequency triazole resistance found In nonculturable Aspergillus fumigatus from lungs of patients with chronic fungal disease.** *Clin Infect Dis* 2011, **52:**1123–1129.

175. Rogers GB, Marsh P, Stressmann AF, Allen CE, Daniels TV, Carroll MP, *et al*: **The exclusion of dead bacterial cells is essential for accurate molecular analysis of clinical samples.** *Clin Microbiol Infect* 2010, **16:**1656–1658.

176. Rogers GB, Stressmann FA, Koller G, Daniels T, Carroll MP, Bruce KD: **Assessing the diagnostic importance of nonviable bacterial cells in respiratory infections.** *Diagn Microbiol Infect Dis* 2008, **62:**133–141.

177. Sauni R, Uitti J, Jauhiainen M, Kreiss K, Sigsgaard T, Verbeek JH: **Remediating buildings damaged by dampness and mould for preventing or reducing respiratory tract symptoms, infections and asthma.** *Cochrane Database Syst Rev* 2011**:**CD007897.

178. Burr ML, Matthews IP, Arthur RA, Watson HL, Gregory CJ, Dunstan FD, *et al*: **Effects on patients with asthma of eradicating visible indoor mould: a randomised controlled trial.** *Thorax* 2007, **62:**767–772.

179. Jarvis JQ, Morey PR: **Allergic respiratory disease and fungal remediation in a building in a subtropical climate.** *Appl Occup Environ Hyg* 2001, **16:**380–388.

180. Patovirta RL, Husman T, Haverinen U, Vahteristo M, Uitti JA, Tukiainen H, *et al*: **The remediation of mold damaged school–a three-year follow-up study on teachers' health.** *Cent Eur J Public Health* 2004, **12:**36–42.

181. Ebbehoj NE, Hansen MO, Sigsgaard T, Larsen L: **Building-related symptoms and molds: a two-step intervention study.** *Indoor Air* 2002, **12:**273–277.

182. Shortt N, Rugkasa J: **"The walls were so damp and cold" fuel poverty and ill health in Northern Ireland: results from a housing intervention.** *Health Place* 2007, **13:**99–110.

183. Howden-Chapman P, Matheson A, Crane J, Viggers H, Cunningham M, Blakely T, *et al*: **Effect of insulating existing houses on health inequality: cluster randomised study in the community.** *BMJ* 2007, **334:**460.

184. Patovirta RL, Meklin T, Nevalainen A, Husman T: **Effects of mould remediation on school teachers' health.** *Int J Environ Health Res* 2004, **14:**415–427.

185. Sudakin DL: **Toxigenic fungi in a water-damaged building: an intervention study.** *Am J Ind Med* 1998, **34:**183–190.

186. Rudblad S, Andersson K, Stridh G, Bodin L, Juto JE: **Slowly decreasing mucosal hyperreactivity years after working in a school with moisture problems.** *Indoor Air* 2002, **12:**138–144.

187. Paba E, Chiominto A, Marcelloni AM, Proietto AR, Sisto R: **Exposure to airborne culturable microorganisms and endotoxin in two Italian poultry slaughterhouses.** *J Occup Environ Hyg* 2014**:** [epub ahead of print].

188. Cafarchia C, Camarda A, Iatta R, Danesi P, Favuzzi V, Di Paola G, *et al*: **Environmental contamination by Aspergillus spp. in laying hen farms and associated health risks for farm workers.** *J Med Microbiol* 2014, **63:**464–470.

189. Rimac D, Macan J, Varnai VM, Vucemilo M, Matkovic K, Prester L, *et al*: **Exposure to poultry dust and health effects in poultry workers: impact of mould and mite allergens.** *Int Arch Occup Environ Health* 2010, **83:**9–19.

190. Rees D, Nelson G, Kielkowski D, Wasserfall C, da Costa A: **Respiratory health and immunological profile of poultry workers.** *S Afr Med J* 1998, **88:**1110–1117.

191. Poole CJ, Wong M: **Allergic bronchopulmonary aspergillosis in garden waste (compost) collectors--occupational implications.** *Occup Med (Lond)* 2013, **63:**517–519.

192. **Inhaled beclomethasone dipropionate in allergic bronchopulmonary aspergillosis. Report to the Research Committee of the British Thoracic Association.** *Br J Dis Chest* 1979, **73**(4)**:**349–356.

193. Agarwal R, Khan A, Aggarwal AN, Saikia B, Gupta D, Chakrabarti A: **Role of inhaled corticosteroids in the management of serological allergic bronchopulmonary aspergillosis (ABPA).** *Intern Med* 2011, **50:**855–860.

194. Barnes PJ, Adcock IM: **Glucocorticoid resistance in inflammatory diseases.** *Lancet* 2009, **373:**1905–1917.

195. Braunstahl GJ, Chen CW, Maykut R, Georgiou P, Peachey G, Bruce J: **The eXpeRience registry: the 'real-world' effectiveness of omalizumab in allergic asthma.** *Respir Med* 2013, **107:**1141–1151.

196. Kudoh S, Keicho N: **Diffuse panbronchiolitis.** *Clin Chest Med* 2012, **33:**297–305.

197. Serisier DJ, Martin ML: **Long-term, low-dose erythromycin in bronchiectasis subjects with frequent infective exacerbations.** *Respir Med* 2011, **105:**946–949.

198. Cameron EJ, McSharry C, Chaudhuri R, Farrow S, Thomson NC: **Long-term macrolide treatment of chronic inflammatory airway diseases: risks, benefits and future developments.** *Clin Exp Allergy* 2012, **42:**1302–1312.

199. Kellett F, Redfern J, Niven RM: **Evaluation of nebulised hypertonic saline (7%) as an adjunct to physiotherapy in patients with stable bronchiectasis.** *Respir Med* 2005, **99:**27–31.

200. Kussek P, Rosario Filho NA, Cat M: **Bronchial hyperresponsiveness to hypertonic saline challenge in children and adolescents.** *J Bras Pneumol* 2006, **32:**195–201.

201. Stevens DA, Schwartz HJ, Lee JY, Moskovitz BL, Jerome DC, Catanzaro A, *et al*: **A randomized trial of itraconazole in allergic bronchopulmonary aspergillosis.** *N Engl J Med* 2000, **342:**756–762.

202. Wark PA, Hensley MJ, Saltos N, Boyle MJ, Toneguzzi RC, Epid GD, *et al*: **Anti-inflammatory effect of itraconazole in stable allergic bronchopulmonary aspergillosis: a randomized controlled trial.** *J Allergy Clin Immunol* 2003, **111:**952–957.

203. Platts-Mills TA, Woodfolk JA: **Trichophyton asthma.** *Chest* 2009, **135:**887–888.

204. Ward GW Jr, Woodfolk JA, Hayden ML, Jackson S, Platts-Mills TA: **Treatment of late-onset asthma with fluconazole.** *J Allergy Clin Immunol* 1999, **104:**541–546.

205. Hurlimann A, Fah J: **Asthma, rhinitis and dermatitis triggered by fungal infection: therapeutic effects of terbinafine.** *Dermatology* 2001, **202:**330–332.

206. Pasqualotto AC, Powell G, Niven R, Denning DW: **The effects of antifungal therapy on severe asthma with fungal sensitization and allergic bronchopulmonary aspergillosis.** *Respirology* 2009, **14:**1121–1127.

207. Agbetile J, Bourne M, Fairs A, Hargadon B, Desai D, Broad C, Morley J, Bradding P, Brightling CE, Green RH, Haldar P, Pashley CH, Pavord ID, Wardlaw AJ: **Effectiveness of voriconazole in the treatment of Aspergillus fumigatus-associated asthma (EVITA3 study).** *J Allergy Clin Immunol* 2013**:** . doi: 10.1016/j.jaci.2013.09.050. [Epub ahead of print].

208. Sermet-Gaudelus I, Lesne-Hulin A, Lenoir G, Singlas E, Berche P, Hennequin C: **Sputum itraconazole concentrations in cystic fibrosis patients.** *Antimicrob Agents Chemother* 2001, **45:**1937–1938.

209. Rodvold KA, Yoo L, George JM: **Penetration of anti-infective agents into pulmonary epithelial lining fluid: focus on antifungal, antitubercular and miscellaneous anti-infective agents.** *Clin Pharmacokinet* 2011, **50:**689–704.

210. Coughlan CA, Chotirmall SH, Renwick J, Hassan T, Low TB, Bergsson G, *et al*: **The effect of Aspergillus fumigatus infection on vitamin D receptor expression in cystic fibrosis.** *Am J Respir Crit Care Med* 2012, **186:**999–1007.

211. Howard SJ, Cerar D, Anderson MJ, Albarrag A, Fisher MC, Pasqualotto AC, *et al*: **Frequency and evolution of Azole resistance in Aspergillus fumigatus associated with treatment failure.** *Emerg Infect Dis* 2009, **15:**1068–1076.

212. Bueid A, Howard SJ, Moore CB, Richardson MD, Harrison E, Bowyer P, *et al*: **Azole antifungal resistance in Aspergillus fumigatus: 2008 and 2009.** *J Antimicrob Chemother* 2010, **65:**2116–2118.

213. Foundation ERSEL: **Pneumonia**. In *European Lung White Book.* 2nd edition. Sheffield; 2003. http://www.erswhitebook.org.

214. Westerink MA, Schroeder HW Jr, Nahm MH: **Immune responses to pneumococcal vaccines in children and adults: rationale for age-specific vaccination.** *Aging Dis* 2012, **3:**51–67.

215. Assaad U, El-Masri I, Porhomayon J, El-Solh AA: **Pneumonia immunization in older adults: review of vaccine effectiveness and strategies.** *Clin Interv Aging* 2012, **7:**453–461.

216. Rozenbaum MH, van Hoek AJ, Fleming D, Trotter CL, Miller E, Edmunds WJ: **Vaccination of risk groups in England using the 13 valent pneumococcal conjugate vaccine: economic analysis.** *Bmj* 2012, **345:**e6879.

217. Vila-Corcoles A, Ochoa-Gondar O: **Pneumococcal vaccination among adults with chronic respiratory diseases: a historical overview.** *Expert Rev Vaccines* 2012, **11:**221–236.

218. White L, Mirrani G, Grover M, Rollason J, Malin A, Suntharalingam J: **Outcomes of Pseudomonas eradication therapy in patients with non-cystic fibrosis bronchiectasis.** *Respir Med* 2012, **106:**356–360.

219. Zelenitsky SA, Rubinstein E, Ariano RE, Zhanel GG: **Integrating pharmacokinetics, pharmacodynamics and MIC distributions to assess changing antimicrobial activity against clinical isolates of Pseudomonas aeruginosa causing infections in Canadian hospitals (CANWARD).** *J Antimicrob Chemother* 2013, **68**(Suppl 1)**:**i67–i72.

220. Stockmann C, Sherwin CM, Zobell JT, Young DC, Waters CD, Spigarelli MG, *et al*: **Optimization of anti-pseudomonal antibiotics for cystic fibrosis pulmonary exacerbations: III: fluoroquinolones.** *Pediatr Pulmonol* 2013, **48:**211–220.

221. Taccetti G, Bianchini E, Cariani L, Buzzetti R, Costantini D, Trevisan F, *et al*: **Early antibiotic treatment for Pseudomonas aeruginosa eradication in patients with cystic fibrosis: a randomised multicentre study comparing two different protocols.** *Thorax* 2012, **67:**853–859.
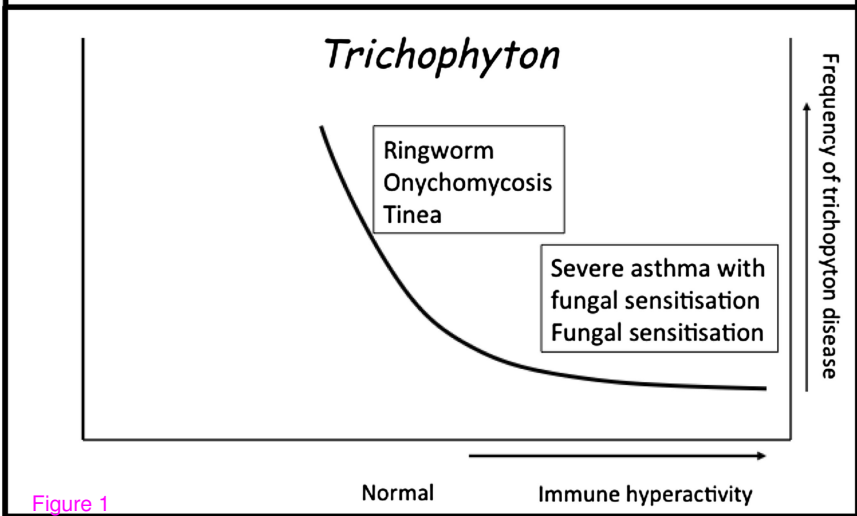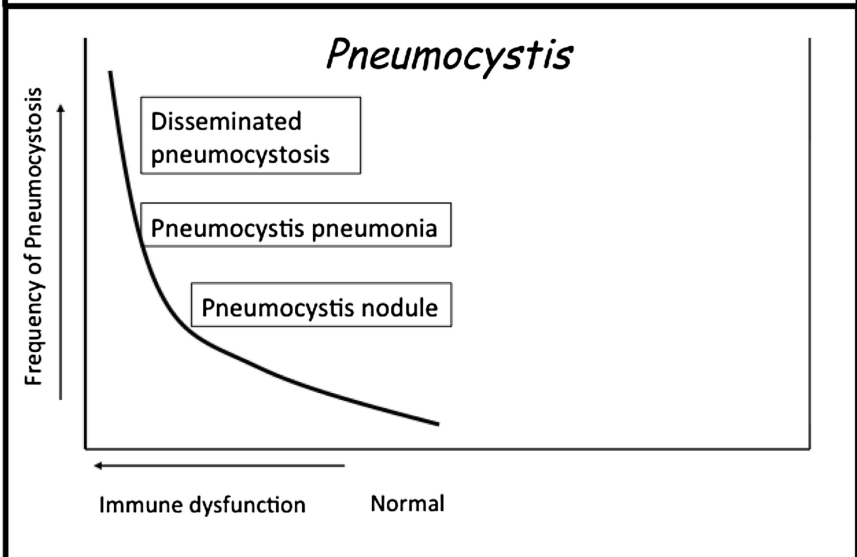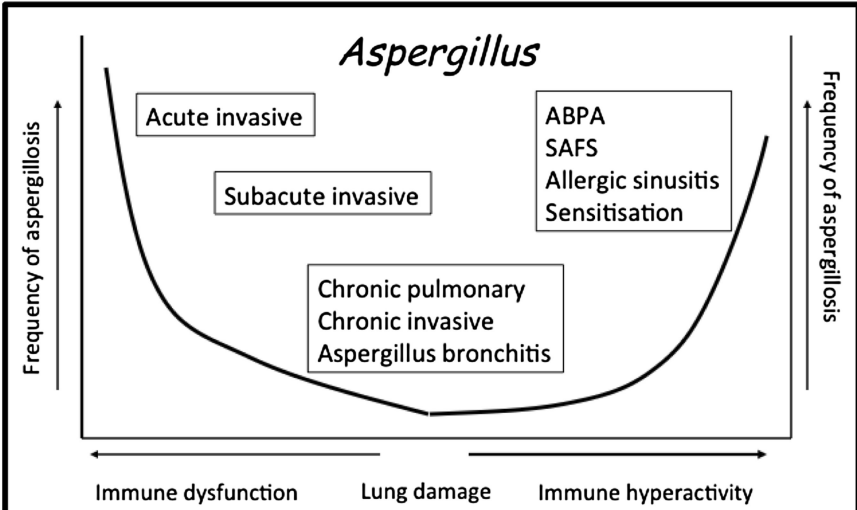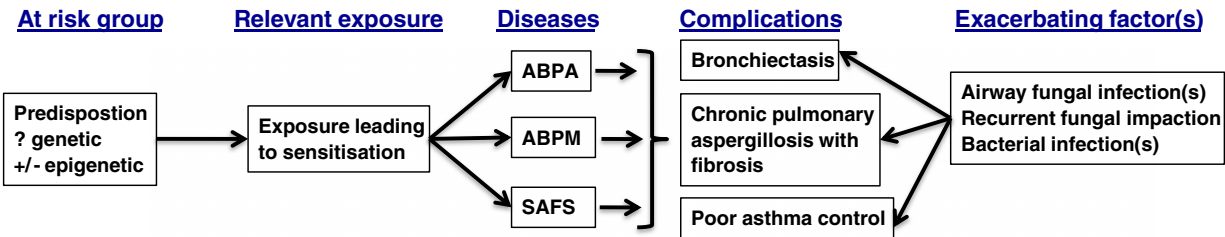
Figure 1

A

| At risk group | Relevant exposure | Diseases | Complications | Exacerbating factor(s) |

At risk group

Predispostion
? genetic
+/- epigenetic

Relevant exposure

Exposure leading to sensitisation

Diseases

ABPA

ABPM

SAFS

Complications

Bronchiectasis

Chronic pulmonary aspergillosis with fibrosis

Poor asthma control

Exacerbating factor(s)

Airway fungal infection(s)
Recurrent fungal impaction
Bacterial infection(s)

B

At risk group

Predispostion
? genetic
+/- epigenetic

Relevant exposure

Specific fungal exposure or infection, ie with viral infection, or skin/mucosal infection

Exposure outcomes

Sensitisation only

Allergic disease ie ABPA, ABPM, SAFS

Bronchiectasis

Chronic pulmonary aspergillosis with fibrosis

Poor asthma control

Exacerbating factor(s)

Airway fungal infection(s)
Recurrent fungal impaction
Bacterial infection(s)

Figure 2