

### REVIEW ARTICLE

## Fungi: the neglected allergenic sources

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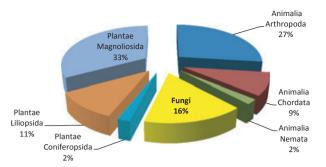
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#### Abstract

Allergic diseases are considered the epidemics of the twentieth century estimated to affect more than 30% of the population in industrialized countries with a still increasing incidence. During the past two decades, the application of molecular biology allowed cloning, production and characterization of hundreds of recombinant allergens. In turn, knowledge about molecular, chemical and biologically relevant allergens contributed to increase our understanding of the mechanisms underlying IgE-mediated type I hypersensitivity reactions. It has been largely demonstrated that fungi are potent sources of allergenic molecules covering a vast variety of molecular structures including enzymes, toxins, cell wall components and phylogenetically highly conserved cross-reactive proteins. Despite the large knowledge accumulated and the compelling evidence for an involvement of fungal allergens in the pathophysiology of allergic diseases, fungi as a prominent source of allergens are still largely neglected in basic research as well as in clinical practice. This review aims to highlight the impact of fungal allergens with focus on asthma and atopic dermatitis.

Allergy is a disease with many faces that can affect different organs like upper and lower respiratory tract, eyes, intestinal tract and the skin. Depending on the affected organ, allergic symptoms manifest as allergic rhinitis (1), allergic asthma (2), IgE-associated atopic dermatitis (3), food allergy (4) or insect venom allergy (5), to mention only the most important ones. The common hallmark of allergic diseases is a switch to the production of allergen-specific IgE raised against normally innocuous environmental allergens (6) that, in special cases, might also cross-react with self-antigens (7, 8). At this asymptomatic stage, the individual is sensitized to a given allergenic source due to the presence of allergen-specific IgE in serum, a condition also called 'atopy'. Detection of allergen-specific IgE is considered as a specific biomarker for the atopic state in clinical practice, which allows in most cases a linkage of a symptom to a particular allergen exposure (9). Measurement of allergen-specific IgE antibodies in serum is normally performed with fully automated devices (10) and used to confirm sensitization to a particular allergen in support of a history-based clinical diagnosis of allergy or a symptom-based suspicion. In sensitized (atopic) individuals, however, re-exposure to the offending allergen induces crosslinking of the high-affinity receptor FccRI-bound allergenspecific IgE on effector cells and, thus, immediate release of anaphylactogenic mediators (11). Although the mechanisms leading to allergic reactions (12, 13) and the sources of exposure are quite well known, our knowledge about the repertoire of molecular structures involved in the pathogenesis of allergic reactions is still rudimentary (14) even if it is well recognized that only a minor fraction of the myriad of proteins to which humans are exposed provokes allergic reactions. Bioinformatics analyses based on structural motifs (15) and BLAST similarity search methods (16) involving 101 602 and 135 850 protein entries deposited in the Swiss-Prot database predict 4093 (4%) and 4768 (3.5%) different potential allergen structures, respectively. Therefore, one can assume that the size of the allergen repertoire involved in eliciting allergic symptoms is in the range of 5000 different structures (14). The modest number of 753 allergenic proteins approved by the World Health Organization and International Union of Immunological Societies (WHO/IUIS) Allergen Nomenclature Subcommittee (www.allergen.org) clearly shows our lack of knowledge in this field. Allergenic structures can be found in every species (Fig. 1), and the number of single allergens characterized is unevenly distributed among the species ranging from 1 for the phylum Cnidaria (sponges and jellyfish) to 252 for the Magnoliopsida trees (flowering plants). However, this is rather due to the number of laboratories working with the different allergenic sources than to the true presence of allergenic structures among these species. Interestingly, the most recent review dealing with nomenclature and structural biology of allergens (17) states 'most major allergens from



Distribution of characterized allergens by tree

**Figure 1** Taxonomic distribution of the 753 allergens officially recognized by the World Health Organization and International Union of Immunological Societies (WHO/IUIS) Allergen Nomenclature Subcommittee (www.allergen.org). The WHO/IUIS allergen nomenclature database has been recently updated (138).

mites, animal dander, pollens, insects, and foods have been cloned, and more than 40 three-dimensional allergen structures are in the Protein Database' with fungal allergens conspicuously absent from this list. Fungal allergens are largely neglected in the field of molecular allergology and also in some reviews dealing with inhalant allergens and their role in allergic airways disease (18) despite the predominant role played by fungi in allergic asthma (19, 20).

#### Allergenic fungi and exposure to fungal allergens

Indoor and outdoor exposure to fungal components, including spores, is a recognized triggering factor for respiratory allergy and asthma (21), as well as for atopic dermatitis (22). As highlighted in a pivotal review (23), among the over 100' 000 fungal species reported (24), only a few hundred have been described as opportunistic pathogens (25) causing human illness through three specific mechanisms: direct infection of the host, elicitation of deregulated immune responses and toxic effects due to secondary metabolites (26). Among these, about 80 mould genera have been shown to induce type I allergies in atopic individuals (23). The most important allergenic fungi belong to the genera *Alternaria*, *Aspergillus* and *Cladosporium* (27), whereas members of the genera *Candida*, *Penicillium*, *Clavularia* and others seem to be, with the exception of the genus *Malassezia* in patients suffering from atopic dermatitis (22, 28), less important as allergenic sources (29).

The exact prevalence of fungal sensitization among the general population is still unknown (27). This is not astonishing as no reference standards for fungal extracts are available to date even if dozens of commercial products are available (30). However, it is well known that the use of commercial extracts from different manufacturers can generate huge differences in the outcome of in vitro or in vivo prevalence studies (31) due to the extreme variability of both content and relative amounts of allergens in commercially available fungal extracts (32). In conclusion, the marked differences in source materials and manufacturing procedures used, the lack of generally accepted potency assays and the great number of potentially allergenic fungal species have hampered any significant progress in fungal extract standardization. The best estimates of fungal sensitization among allergic individuals come from a large skin prick test (SPT) survey on a cohort of subjects with respiratory diseases conducted with extracts from Alternaria, Aspergillus, Candida, Cladosporium, Penicillium, Saccharomyces and Trichophyton and indicate a prevalence of sensitization around 19% (29). The ranges of prevalence of fungal sensitization for the general population, atopic and asthmatic subjects, and mould-sensitized patients, not claimed to be exhaustive, are reported in Table 1.

#### Clinical manifestations of fungal hypersensitivity

The clinical spectrum of hypersensitivity reactions elicited by fungi is very broad and includes, besides IgE-mediated type I allergy, reactions of types II, III and IV according to the old definition of Coombs and Gell (33, 34). Although the classification into types I to VI is widely used in clinical practice, it should be mentioned that the reality is more complex because frequently several mechanisms operate together in the pathogenesis of hypersensitivity reactions, and this is especially true for reactions to fungi. A brief classification of the different types of hypersensitivity reactions with the mechanisms

Table 1 Prev	alence of mould	allerav induced b	ov different funga	I species in% of the	e respective p	oopulations investigated

Genus	General population	Atopics*	Asthmatics	Mould-allergic individuals†
Alternaria	3.6–12.6 (20, 29)	3–14.6 (31, 139)	13.5–14.6 (140, 141)	66.1 (29)
Aspergillus	2.4 (29)	15-27.6 (22, 140)	5–21.3 (140, 141)	12.6 (29)
Candida	8.5 (29)	28,9 (22)	23.1 (141)	44.3 (29)
Cladosporium	2.5-2.9 (20, 46)	3–18.2 (31, 139)	15.9 (141)	13.1 (29)
Penicillium	1.5 (29)	7.3–13.1 (22, 139)	33 (142)	33 (142)
Trichophyton	1.9 (29)	ND	NA (29)*	10.2 (29)

\*Individuals suffering from allergen-specific IgE-mediated sensitization against any allergenic source.

†Individuals sensitized to at least one mould. The prevalence of sensitization to single moulds in the subpopulation of subjects with multiple fungal sensitizations might reach prevalences surpassing 80% for *Alternaria* (92.2%), *Aspergillus* (83.1%), *Candida* (89.6%) and *Cladosporium* (84.4%) (29).

involved in their pathogenesis is reported in Table 2. As this review focuses on IgE-mediated type I allergic reactions, hypersensitivity reactions of types II, III and IV will not be further discussed in detail.

Fungal type I allergy is induced by a large number of fungal genera, the most important ones belonging to the ascomycota followed by basidiomycota and zygomycota (23). Clinically, the IgE-mediated sensitization to fungal allergens can manifest as allergic rhinitis and rhinosinusitis (35), allergic asthma (36) and atopic dermatitis (37).

#### Allergic rhinitis and rhinosinusitis

Allergic rhinitis affects up to 40% of the population and results in nasal itching, congestion, sneezing and clear rhinorrhea. Allergic rhinitis causes extranasal adverse effects including decreased quality of life, decreased sleep quality and obstructive sleep apnoea (38). However, epidemiologic studies have failed to demonstrate a direct relationship between fungal allergy and allergic rhinitis via either outdoor or indoor exposure. Fungal allergy is clearly linked to a subset of chronic rhinosinusitis (CRS) known as allergic fungal rhinosinusitis (AFRS) (39). In the patient's mucus, fungal hyphae are detectable and patients show coetaneous hypersensitivity to specific fungal allergens along with specific IgE and IgG antibodies against the sensitizing fungus and an increased total serum IgE level (40). Immunologically, allergic fungal rhinosinusitis is a mixed type I, type III and type IV allergic reaction.

#### Allergic asthma

There is compelling evidence that fungal allergy is associated with severe asthma (41). Although the precise prevalence of fugal sensitivity is unclear, recent estimates indicate that 24.6 million people in the United States suffer from asthma (42). Depending on the definition, about 10–20% of these

Table 2 Classification of hypersensitivity reactions\*

patients might be classified as subjects suffering from severe asthma, and in this group, 30–70% can be expected to be sensitized to at least one fungal species (Table 1). Extrapolating this figure to the industrialized countries, we have to assume that several millions of asthmatic patients are affected by fungal allergy (27). However, with exception of special cases such as workplace exposure or allergic bronchopulmonary aspergillosis (ABPA), which are well documented, the contribution of fungal sensitization to the severity of asthma remains to be investigated.

# Allergic bronchopulmonary aspergillosis and related conditions

ABPA is commonly caused by Aspergillus fumigatus, a ubiquitous mould frequently found indoors and outdoors. The syndrome is characterized by exacerbations of asthma, recurrent transient chest radiographic infiltrates, peripheral and pulmonary eosinophilia, and development of bronchiectasis (43). Although originally considered a rarity (44), ABPA is currently recognized as a serious disease affecting approximately 3% of asthmatic patients (41) and up to 10% of patients with cystic fibrosis (45). However, in asthmatic and CF patients sensitized to A. fumigatus, the incidence of ABPA might be much higher ranging from 15 to 35% (46, 47). Immunologically, ABPA is a mixed type I, type III and type IV hypersensitivity lung disease induced by bronchial colonization with A. fumigatus causing symptoms ranging from asthma exacerbation to fatal destruction of the lung (48). ABPA as a syndrome suspected from clinical signs is not trivial to be diagnosed, and several criteria that are, however, not specific for the disease have been suggested (45, 49, 50). Therefore, serological findings showing sensitization to A. fumigatus are essential diagnostic tools to confirm or exclude the disease. ABPA is a disease with a high risk of the development of irreversible end-stage fibrosis, and therefore, it is recommended to rule out sensitization to A. fumigatus in

Category†	Humoral response	Soluble mediators	Time course	Cellular response	Clinical examples	Fungal diseases
Туре I	IgE	Histamine, leukotrienes	Minutes	Smooth muscle constriction, eosinophil infiltration	Rhinitis, allergic asthma (143)	Allergic rhinitis (39) Allergic asthma (42, 48) ABPA (133–137) ABPM (23)
Type II	lgG, lgM	Complement	1–24 h	Neutrophil activation and lysis of target cells	Autoimmunity(144)	Unknown
Type III	lgG, lgM	Complement	1–24 h	Infiltration and activation of granulocytes	Rheumatoid arthritis (145)	Hypersensitivity pneumonitis‡ Aspergilloma (137)
Type IV	T cells	Lymphokines	2–3 days	T-cell and macrophage activation	Tuberculosis, contact dermatitis (146, 147)	ABPA (133–137) Hypersensitivity pneumonitis (135, 136)

\*Modified from references (33) and (34).

\*Most of the IgE-associated fungal diseases are mixed forms involving combinations of types I, III and IV hypersensitivity reactions (23). \*Also termed extrinsic allergic alveolitis (137). all asthmatic patients (51). In this regard, recombinant allergens might contribute to a more reliable diagnosis of ABPA.

Other fungi, including *Candida, Penicillium* and *Curvularia* species, are occasionally responsible for a similar syndrome termed 'allergic bronchopulmonary mycosis' (ABPM) (52). The endotype classification of asthma syndromes proposed recently (53) also include ABPM. Like ABPA, the characteristics of ABPM include severe asthma, blood and pulmonary eosinophilia, marked increased levels of total and allergenspecific IgE, bronchiectasis and mould colonization of the airways. The term 'severe asthma associated with fungal sensitivity' (SAFS) has been introduced to illustrate the high rate of fungal sensitivity in patients with severe asthma (54). Because of the ambiguity in diagnostic criteria, SAFS is currently more a diagnosis by exclusion than a diagnosis of a specific disease entity (55).

#### Atopic dermatitis

Atopic dermatitis (AD) is a chronic relapsing, highly pruritic inflammation of the skin with a worldwide prevalence of 10-20% in children and of 1-3% in adults (22, 56). The pathophysiology of AD, also called atopic eczema, is complex and not fully understood (57). Recently, Malassezia sympodialis, a lipophilic yeast colonizing the skin of both AD and healthy individuals, has been shown to induce IgE-mediated sensitization exclusively in patients suffering from AD (22). The main reason for this specific sensitization may be the disrupted skin barrier facilitating allergen uptake, which may contribute to the perpetuation of the disease (58). Of special interest in this regard are cross-reactivity reactions between M. sympodialis allergens sharing a high degree of sequence identity to human proteins (8, 28). It has been convincingly shown, both, in vitro and in vivo that Mala s 11, the M. sympodialis manganese-dependant superoxide dismutase, sharing 50% sequence homology with the human enzyme (59), can elicit strong humoral- and T-cell-mediated immune reactions in a subset of AD patients (60). These reactions have been traced back to structural similarities between the two proteins (59) and studied in detail at the Tcell level (61). However, the phenomenon of autoreactivity to human proteins in AD patients is not limited to Mala s 11 and human superoxide dismutase as recently shown in studies investigating Mala s 13 and human thioredoxin (62, 63) and can be extended to a whole array of human proteins (64). whether sensitization to other fungi, except Saccharomyces cerevisiae, which shows a significant correlation between a positive skin prick test and AD (65), is involved in the pathogenesis of the disease remains to be determined.

#### **Fungal allergens**

The list of fungal allergens officially approved by the Nomenclature Subcommittee of the International Union of Immunological Societies (IUIS; www.allergen.org) spans 105 iso-allergens and variants from 25 fungal species belonging to the Ascomycota and Basidiomycota phyla (20). However, the number of fungal proteins able to elicit type I hypersensitivity

reactions described in the literature is much longer, even if many of these allergens are poorly characterized. A recent catalogue of the fungal allergens described (23) lists 174 allergens for the genus Ascomycota and 30 for the genus Basidiomycota. However, this list does not include many fungal allergens, which have been only partially characterized in terms of primary sequence. For example, it has been shown by high-throughput screening technology that A. fumigatus, perhaps the most important allergenic mould, is able to produce at least eighty-one different IgE-binding proteins (66). The same approach applied to phage surface display libraries of Cladosporium herbarum, Coprinus comatus and Malassezia furfur yielded at last 28, 37 and 27 different clones, respectively, displaying IgE-binding proteins (67). These few examples clearly show that the repertoire of fungal allergens is far from being completely elucidated. Besides species-specific allergens such as Asp f 1 and Alt a 1, which are limited to genera or species (68), databases of allergen sequences compiled and used to search fungal proteomes revealed that some highly homologous allergen orthologue classes and allergen epitopes are ubiquitous in all fungi (69). It seems likely that many of these protein orthologues are potential allergens or at last capable of cross-reacting as proteins showing a high degree of sequence homology are likely cross-reactive (70). Cross-reactivity between homologous fungal allergens has been demonstrated in many cases between phylogenetically close (71, 72) and even distant species such as Candida boidiini and A. fumigatus (73). Some examples of major and cross-reactive fungal allergens are given in Table 3. Although the clinical relevance of crossreactivity between fungal allergens remains to be investigated in more detail (7), the phenomenon is well understood at a scientific level. The availability of high-resolution threedimensional structures of eight fungal allergens (74-81) allowed researchers to understand in details the structural basis of cross-reactivity (70), to homology model unsolved structures of fungal allergens (59, 82-84) and to test the correctness of the hypotheses by site-directed mutagenesis and immunological investigations (59,85). Moreover, serologic studies involving recombinant A. fumigatus allergens contributed to corroborate a clinical diagnosis of ABPA by the discovery of disease-specific allergens (86, 87). In contrast to secreted allergens, which are recognized by serum IgE of A. fumigatus-sensitized individuals with or without ABPA, some nonsecreted allergens are predominantly recognized by serum IgE of ABPA patients (45, 86-89). This differential immunological response is probably due to the fact that ABPA patients have or had the fungus growing in the lung (90). Therefore, as a result of fungal damage due to cellular defence mechanisms, they become more strongly exposed to nonsecreted proteins than A. fumigatus-allergic individuals, which mainly recognize environmental fungal allergens (91).

# The diagnosis of fungal allergy: an unsolved medical need

As already mentioned and confirmed in a recent study supported by the European community (GA<sup>2</sup>LEN (92)), the

 Table 3
 Major fungal allergens with or without cross-reactivity

Allergen	Genus	Accession Number	MW (kDa)	Biological Function	Cross-reactive with*	Ref.
Alt a 1	Alternaria alternata	U82633	30	Unknown	NF	(69)
Asp f 1	Aspergillus fumigatus	M83781	18	Ribotoxin	NF	(68, 69, 148)
Asp f 3	A. fumigatus	U58050	19	Peroxisomal protein	Candida boidiini	(73)
Asp f 11	A. fumigatus	AJ006689	24	Cyclophylin	Asp f 27, Mala s 6, Cyclopylins of Candida albicans, Saccharomyces cerevisiae, Homo sapiens	(75, 149)
Asp f 29	A. fumigatus	AJ937745	13	Thioredoxin	Asp f 28, Mala s 13, <i>H. sapiens</i> thioredoxin	(72, 76)
Asp f 34	A. fumigatus	AM496018	20	Phi A cell wall protein	NF	(150)
Cla h 8	Cladosporium herbarum	AJ181916	28	Mannitol dehydrogenase	Alt a 8	(151)
Pen o 18	Penicillium oxalicum	AF243425	34	Vacuolar serine protease	Cla h 18, Asp f 18	(152)
Mals s 6	Malassezia sympodialis	AJ011956	17	Cyclophilin	Asp f 11, Asp f 27, Asp f 28, <i>H. sapiens</i> cyclophilin	(75)
Mala s 11	M. sympodialis	AJ548421	23	MnSOD	Asp f 6, <i>H. sapiens</i> MnSOD	(60, 74, 85)

\*NF: not found.

These allergens are considered species specific and unique (69). Many phylogenetically highly conserved allergens react also with their human counterpart.

incidence of fungal sensitization is high (Table 1) and clinically relevant. However, the problems related to the in vitro and in vivo diagnosis of fungal and other allergies are far from being solved (93). Of course any in vivo diagnosis of allergy based on skin tests as well as any in vitro diagnosis of allergy based on the determination of allergen-specific IgE depends on the quality of the material used for testing. Although standardized extracts approved by the FDA are available for some allergenic sources (94, 95), no fungal extracts have been approved to date. This is not astonishing because, in contrast to pollens or insect venoms, which represent allergenic sources more or less 'standardized' by nature as a feature of their biological function, fungi as allergenic source are extremely complex. Problems encountered in the standardization of fungal extracts derive from different patterns of allergens produced by different clinical isolates (96), batch to batch variations (97), time-dependent liberation of IgE-binding components during fungal growth (98), instability of the extracts due to protease content (99), culture conditions and medium used to grow the fungus and, of course, the extraction procedures (100). Moreover, fungi produce cell-bound, secreted and intracellular allergens (101), making it impossible to produce consistent extracts containing the three types of allergens in a single run. Therefore, even commercial extracts for skin tests provided by different suppliers deliver discrepant results (102). In clinical routine, fungal sensitization is normally assessed in vitro by determination of serum IgE levels with ImmunoCAPs, still considered the 'gold standard' for the in vitro diagnosis of allergy (103). Unfortunately, the ImmunoCAP manufacturer does not provide the extracts used to produce the in vitro test system as skin test solution, hampering a direct comparison of the skin test reactivity of a commercial extract with the serum IgE levels determined with the corresponding ImmunoCAP. This would be extremely important because (fungal) extracts con-

tain the so-called cross-reactive carbohydrate determinants to which about 20% or more of the sensitized patients generate specific antiglycan IgE (104). Even though antibody-binding glycoproteins are widespread in many extracts and therefore detected by in vitro diagnostic tests, cross-reactive carbohydrate determinants do not appear to cause clinical symptoms in most, if not all, patients and can thus be considered as clinically irrelevant allergens (104). In fact, comparisons of skin test and serology obtained with recombinant allergens produced in E. coli, and thus lacking post-translational modifications, correlate fairly well (105) in contrast to those obtained with fungal extracts (106). Moreover, the clinical history that represents one of the most important diagnostic criteria for an allergy is difficult to reconstruct for patients with a fungal allergy. In contrast to other allergies such as seasonal pollen-derived allergies, most patients sensitized to fungi are not aware of the source of exposure and can only report that the symptoms are more or less perennial. Technological developments in allergen cloning and microarrays (107-110), together with the proved superior diagnostic performance of recombinant allergens compared with extracts (111-113), might contribute to a more specific and sensitive component-resolved diagnosis of allergy (114-116) in the future. However, due to the high number of different allergens produced by fungi, it is unlikely that a solution for this urgent medical need will be reached in the near future, and the first recombinant fungal allergens (Alt a 1, Asp f 1, Asp f 2, Asp f 3 and Asp f 4) immobilized in ImmunoCAPs are now commercially available from Thermo scientific (www. phadia.com).

#### The treatment of fungal allergy

As for all other allergies, avoiding exposure is still the best treatment for these diseases. Because fungi are ubiquitous in our environment, a complete avoidance of exposure is not feasible. However, reduction in asthma morbidity following interventions for improving indoor air quality and remediation of moisture incursion have been demonstrated (117).

Of course, allergen-specific immunotherapy is currently the only treatment capable of curing allergic diseases (118) and has been used in clinical practice for more than hundred years (119). A limited number of controlled immunotherapy trials with *Alternaria alternata* and *C. herbarum* extracts have indicated some clinical benefit (120); however, large-scale, double-blind, placebo-controlled studies of fungal allergen-specific immunotherapy are lacking. In general, immunotherapy for fungal allergy is not recommended mainly because of the lack of standardized therapeutic reagents (121).

As the majority of the patients suffering from fungal sensitization are severe asthmatics, inhaled corticosteroids and frequent courses of oral corticosteroids are used to control the asthma (122, 123) and contribute to alleviate the allergic symptoms (124). Severe conditions such as ABPA, ABPM or SAFS exacerbations are best treated with oral steroids over 3-6 weeks (123) corroborated by inhaled corticosteroids. Although there are conflicting data concerning the clinical utility of inhaled corticosteroids in reducing exacerbation frequency, they are an important therapeutic intervention to control the worst symptoms of underlying asthma (125, 126), but with the well-known adverse side-effects (127). An alternative or adjunctive strategy in the treatment of these diseases is to reduce or clear the lung of fungal colonization by antifungal agents (128). Although some placebo-controlled randomized studies demonstrated the benefit of itraconazole therapy for ABPA (129, 130), the adjuvant role of antifungal treatment in severe forms of fungal allergy should be investigated in more detail with special attention to the effects on corticosteroid dose, frequency of exacerbations, quality of life, immunological changes and side-effects.

#### Other diseases associated with fungal exposure

Fungi play an important role also in other important diseases associated with fungal exposure, although not primarily IgE mediated. As this review is focused on fungal allergy, these

#### References

- Greiner AN, Hellings PW, Rotiroti G, Scadding GK. Allergic rhinitis. *Lancet* 2011;**378**:2112–2122.
- Sullivan SD, Turk F. An evaluation of the cost-effectivness of omalizumab for the treatment of severe allergic asthma. *Allergy* 2008;63:670–684.
- Bieber T, Cork M, Reitamo S. Atopic dermatitis: a candidate for disease-modifying strategy. *Allergy* 2012;67:969–975.
- Berin MC, Sicherer S. Food allergy: mechanisms and therapeutics. *Curr Opin Immunol* 2011;23:794–800.
- Müller UR. Insect venoms. Chem Immunol Allergy 2010;95:141–156.

- Galli SJ, Tsai M, Piliponsky AM. The development of allergic inflammation. *Nature* 2008;**254**:445–454.
- Zeller S, Glaser AG, Vilhelmsson M, Rhyner C, Crameri R. Cross-reactivity among fungal allergens: a clinically relevant phenomenon? *Mycoses* 2009;**52**:99–106.
- Crameri R. Immunoglobulin E-binding autoantigens: biochemical characterization and clinical relevance. *Clin Exp Allergy* 2012;42:343–351.
- Hamilton RG, Donald MacGlashan WW Jr, Saini SS. IgE antibody-specific activity in human allergic disease. *Immunol Res* 2010;47:273–284.
- Hamilton RG, Adkinson FN Jr. In vitro assays for the diagnosis of IgE-mediated disorders. J Allergy Clin Immunol 2004;114:213–215.
- Peavy RD, Metcalfe DD. Understanding the mechanisms of anaphylaxis. *Curr Opin Allergy Clin Immunol* 2008;83:305–310.
- Bousquet J, Anto J, Auffray C, Akdis M, Cambon-Thomsen A, Keil T et al. MeDALL (Mechanisms of the Development of ALLergy): an integrated approach from phenotypes to systems medicine. *Allergy* 2011;66:596–604.
- Boyce JA, Bochner B, Finkelman FD, Rothenberg ME. Advances in mechanisms

diseases are only briefly mentioned here. The most dangerous diseases caused by fungi are invasive mycoses (131, 132). Most opportunistic mycoses occur in individuals with congenital or acquired immunodeficiency; morbidity and mortality rates are high, and prevention, diagnosis and treatment of these infections remain difficult (133). Less harmful but widespread in the population are fungal infections of the skin. Superficial mycoses are characterized by invasion restricted to the stratus corneum and therefore usually not associated with inflammatory or immune responses of the host (134). Other diseases caused by fungi partially are related to exposure at the workplace (135). Included in this disease group are hypersensitivity pneumonitis also called extrinsic allergic alveolitis (136), farmer's lung, bagassosis and mushroom worker's lung, which result from occupational exposure to thermophilic actinomycetes present in hay, bagasse and mushroom compost, respectively (137).

### Conclusions

The impact of fungal allergy on human health, especially in patients suffering from asthma or cystic fibrosis, and an emerging role of Malassezia sensitization in the exacerbation of atopic dermatitis have been clearly demonstrated during the past years. There is no doubt that fungi are involved in many allergic disorders and, as best documented example, in ABPA.

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#### **Conflicts of interest**

The authors declare no conflicts of interest.

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of asthma, allergy, and immunology in 2011. J Allergy Clin Immunol 2012;**129**:335-341.

- Crameri R. Allergy diagnosis, allergen repertoires, and their implication in allergenspecific immunotherapy. *Immunol Allergy Clin North Am* 2006;26:179–189.
- Stadler MB, Stadler BM. Allergenicity prediction by protein sequence. *FASEB J* 2003;17:1141–1143.
- Li KB, Issac P, Krishnan A. Predicting allergenic proteins using wavelet transform. *Bioinformatics* 2004;20:2572–2578.
- Chapman MD, Pomés A, Breiteneder H, Ferreira F. Nomenclature and structural biology of allergens. J Allergy Clin Immunol 2007;119:414–420.
- Custovic A, Simpson A. The role of inhalant allergens in allergic airways disease. J Invest Allergol Clin Immunol 2012;22:393– 401.
- Kennedy JL, Heymann PW, Platts-Mills TAE. The role of allergy in severe asthma. *Clin Exp Allergy* 2012;42:659–669.
- Agarwal R, Gupta D. Severe asthma and fungi: current evidence. *Med Mycol* 2011;49 (Suppl 1):S150–S157.
- Green BJ, Tovey ER, Sercombe JK, Blachere FM, Beezhold DH, Schmechel D. Airborne fungal fragments and allergenicity. *Med Mycol* 2006;44(Suppl 1):S245–S255.
- Gaitanis G, Magiatis P, Hantschke M, Bassukas ID, Velegraki A. The *Malassezia* genus in skin and systemic diseases. *Clin Microbiol Rev* 2012;25:106–141.
- Simon-Nobbe B, Denk U, Pöll V, Rid R, Breitenbach M. The spectrum of fungal allergy. *Int Arch Allergy Immunol* 2008;145:58–86.
- 24. Prillinger H, Lopandic K, Schweigkofler W, Deak R, Aarts HJ, Bauer R et al. Phylogeny and systematics of the fungi with special reference to the Ascomycota and Basidiomycota. *Chem Immunol* 2002;81:207–295.
- Horner WE, Helbling A, Salvaggio EJ, Lehrer SB. Fungal allergens. *Clin Microbiol Rev* 1995;8:161–179.
- Bush RK, Portneoy JA, Saxon A, Terr AI, Wood RA. The medical effects of mold exposure. J Allergy Clin Immunol 2006;117:326–333.
- Crameri R, Weichel M, Flückiger S, Glaser AG, Rhyner C. Fungal allergies: a yet unsolved problem. *Chem Immunol Allergy* 2006;91:121–133.
- Schmid-Grendelmeier P, Scheynius A, Crameri R. The role of sensitization to Malassezia sympodialis in atopic eczema. Chem Immunol Allergy 2006;91:98–109.
- Mari A, Schneider P, Wally V, Breitenbach M, Simon-Nobbe B. Sensitization to fungi: epidemiology, comparative skin tests, and

IgE reactivity of fungal extracts. *Clin Exp Allergy* 2003;**33**:1429–1438.

- Esch RE. Manufacturing and standardizing fungal allergen products. J Allergy Clin Immunol 2004;113:210–215.
- 31. D'Amato G, Chatzigeorgiou G, Corsico R, Gioulekas D, Jäger L, Jäger S et al. Evaluation of the prevalence of skin prick test positivity to *Alternaria* and *Cladosporium* in patients with suspected respiratory allergy. A European multicenter study promoted by the Subcommittee on Aerobiology and Environmental Aspects of Inhalant Allergens of the European Academy of Allergology and Clinical Immunology. *Allergy* 1997;**52**:711–716.
- 32. Vailes L, Sridhara S, Cromwell O, Weber B, Breitenbach M, Chapman M. Quantitation of the major fungal allergens, Alt a 1 and Asp f 1, in commercial allergenic products. *J Allergy Clin Immunol* 2001;**107**:641–646.
- Coombs RRA, Gell PGH. Clinical Aspects of Immunology. Oxford: Blackwell Scientific Publications, 1963.
- Descotes J, Choquet-Kastylevsky G. Gell and Coomb's classification: is it still valid? *Toxicol* 2001;158:43–49.
- 35. Bousquet J, Schünemann HJ, Samolinski B, Demoly P, Baena-Cagnani CE, Bachert C et al. Allergic rhinitis and its impact on asthma (ARIA): achievements in 10 years and future needs. J Allergy Clin Immunol 2012;130:1049–1062.
- Holgate ST. Innate and adaptive immune responses in asthma. *Nat Med* 2012;18:673– 683.
- Casagrande BF, Flückiger S, Linder MT, Johansson C, Scheynius A, Crameri R et al. Sensitization to the yeast *Malassezia* sympodialis is specific for extrinsic and intrinsic eczema. J Invest Dermatol 2006;**126**:2414–2421.
- Foreman A, Boase S, Psaltis A, Wormald PJ. Role of bacterial and fungal biofilms in chronic rhinosinusitis. *Curr Allergy Asthma Rep* 2012;12:127–135.
- Hamilos DL. Chronic rhinosinusitis: epidemiology and medical management. J Allergy Clin Immunol 2011;128:93–707.
- Kurup VP. Fungal Allergy. In: Arora N editor. *Handbook of Fungal Biotechnology*. New York: Dekker, 2003:515–525.
- Knutsen AP, Bush RK, Demain JG, Denning DW, Dixit A, Fairs A et al. Fungi and allergic lower respiratory tract diseases. *J Allergy Clin Immunol* 2012;129:280–291.
- Akinbami LJ. Asthma prevalence, health care use, and mortality: United States 2005–2009. *Natl Health Stat Report* 2011;**32**:1–16.
- Patterson K, Strek ME. Allergic bronchopulmonary aspergillosis. Proc Am Thorac Soc 2010;7:237–244.

- Slavin RK, Stanczyk DJ, Lonigro AJ, Brown GS. Allergic bronchopulmonary aspergillosis: a North American rarity. *Am* J Med 1969;47:306–313.
- 45. Stevens DA, Moss RB, Kurup VP, Knutsen AP, Greenberger P, Judson MA et al. Allergic bronchopulmonary aspergillosis in cystic fibrosis: Cystic Fibrosis Foundation Consensus Conference. *Clin Infect Dis* 2003;**37**(Suppl):S225–S264.
- Basica JE, Graves TS, Baz MN. Allergic bronchopulmonary aspergillosis in corticoid dependent asthmatics. J Allergy Clin Immunol 1981;68:98–102.
- Laufer P, Fink JN, Bruns WT, Unger GF, Kalbfleisch JH, Greenberger PA et al. Allergic bronchopulmonary aspergillosis in cystic fibrosis. J Allergy Clin Immunol 1984;73:44–48.
- Denning DW, Pleuvry A, Cole DC. Global burden of allergic bronchopulmonary aspergillosis with asthma and its complication chronic pulmonary aspergillosis in adults. *Med Mycol* 2013;51:361–370.
- Greenberger PA, Miller TP, Roberts M, Smith LL. Allergic bronchopulmonary aspergillosis in patients with and without bronchiectasis. *Ann Allergy* 1993;70:333– 338.
- Hafen GM, Hartl D, Regamey N, Casaulta C, Latzin P. Allergic bronchopulmonary aspergillosis: the hunt for a diagnostic serological marker in cystic fibrosis patients. *Expert Rev Mol Diagn* 2009;9:157–164.
- Greenberger PA, Patterson R. Allergic bronchopulmonary aspergillosis and the evaluation of the patient with asthma. J Allergy Clin Immunol 1988;81:645–650.
- 52. Ogawa H, Fujimura M, Takeuchi Y, Makimura K, Satoh K. The definitive diagnostic process and successful treatment for ABPM caused by *Schizophyllum commune*: a report of two cases. *Allergol Int* 2012;61:163–169.
- 53. Lötvall J, Akdis CA, Bacharier LB, Bjermer L, Casale TB, Custovic A et al. Asthma endotypes: a new approach to classification of disease entities within the asthma syndrome. J Allergy Clin Immunol 2011;127:355–3601.
- Denning DW, O'Driscoll BR, Hogaboam CM, Bowyer P, Niven RM. The link between fungi and severe asthma: a summary of the evidence. *Eur Respir J* 2006;27:615–625.
- Agarwal R. Severe asthma with fungal sensitization. *Curr Allergy Asthma Rep* 2011;11:403–413.
- Simon D, Kernland Lang K. Atopic dermatitis: from new pathogenic insights toward a barrier-restoring and antiinflammatory therapy. *Curr Opin Pediatr* 2011;23:647–652.

- Novak N, Simon D. Atopic dermatitis from new pathophysiologic insights to individualized therapy. *Allergy* 2011;66:830–839.
- Wolf R, Wolf D. Abnormal epidermal barrier in the pathogenesis of atopic dermatitis. *Clin Dermatol* 2012;**30**:329–334.
- Vilhelmsson M, Glaser AG, Martinez DB, Schmidt M, Johansson C, Rhyner C et al. Mutational analysis of amino acid residues involved in IgE-binding to the *Malassezia* sympodialis allergen Mala s 11. Mol Immunol 2008;46:294–303.
- Schmid-Grendelmeier P, Flückiger S, Disch R, Trautmann A, Wüthrich B, Blaser K et al. IgE-mediated and T cell-mediated autoimmunity against manganese superoxide dismutase in atopic dermatitis. *J Allergy Clin Immunol* 2005;115:1068– 1075.
- 61. Vilhelmsson M, Johansson C, Jacobsson-Ekman G, Crameri R, Zargari A, Scheynius A. The *Malassezia sympodialis* allergen Mala s 11 induces human dendritic cell maturation, in contrast to its human homologue manganese superoxide dismutase. *Int Arch Allergy Immunol* 2007;**143**:155–162.
- 62. Balaji H, Heratizadeh A, Wichmann K, Niebuhr M, Crameri R, Scheynius A et al. *Malassezia sympodialis* thioredoxin-specific T cells are highly cross-reactive to human thioredoxin in atopic dermatitis. J Allergy Clin Immunol 2011;**128**:e4.
- Hradetzky S, Roesner LM, Heratizadeh A, Crameri R, Scheynius A, Werfel T. Cytokine responses induced by the human autoallergen thioredoxin. *Exp Dermatol* 2013;20:E3.
- Zeller S, Rhyner C, Meyer N, Schmid-Grendelmeier P, Akdis CA, Crameri R. Exploring the repertoire of IgE-binding self-antigens associated with atopic eczema. *J Allergy Clin Immunol* 2009;**124**:278–285.
- Kortekangas-Savolainen O, Lammintausta K, Kalimo K. Skin prick test reactions to brewer's yeast (*Saccharomyces cerevisiae*) in adult atopic dermatitis patients. *Allergy* 1993;48:147–150.
- Kodzius R, Rhyner C, Konthur Z, Buczek D, Lehrach H, Walter G et al. Rapid identification of allergen-encoding cDNA clones by phage display and high-density arrays. *Comb Chem High Throughput Screen* 2003;6:147–154.
- Crameri R, Kodzius R, Konthur Z, Lehrach H, Blaser K, Walter G. Tapping allergen repertoires by advanced cloning technologies. *Int Arch Allergy Immunol* 2001;**124**:43–47.
- Kao R, Martínez-Ruiz A, Martínez del Pozo A, Crameri R, Davies J. Mitogillin and related fungal ribotoxins. *Methods Enzymol* 2001;341:324–335.

- Bowyer P, Fraczek M, Denning DW. Comparative genomics of fungal allergens and epitopes shows widespread distribution of closely related allergen and epitope orthologues. *BMC Genomics* 2006;7:251.
- Aalberse RC, Crameri R. IgE-binding epitopes: a reappraisal. *Allergy* 2011;66:1261– 1274.
- Soeria-Atmadja D, Onnel A, Borga A. IgE sensitization to fungi mirror fungal phylogenetic systematic. J Allergy Clin Immunol 2010;125:1379–1386.
- Glaser AG, Menz G, Kirsch AI, Zeller S, Crameri R, Rhyner C. Auto- and crossreactivity to thioredoxin allergens in allergic bronchopulmonary aspergillosis. *Allergy* 2008;63:1617–1623.
- Hemmann S, Blaser K, Crameri R. Allergens of Aspergillus fumigatus and Candida boidiini share IgE-binding epitopes. Am J Respir Crit Care Med 1997;156:1956–1962.
- 74. Flückiger S, Mittl PR, Scapozza L, Fijten H, Folkers G, Grütter MG et al. Comparison of the crystal structures of the human manganese superoxide dismutase and the homologous *Aspergillus fumigatus* allergen at 2- Å resolution. *J Immunol* 2002;**168**:1267–1272.
- 75. Glaser AG, Limacher A, Flückiger S, Scheynius A, Scapozza L, Crameri R. Analysis of the cross-reactivity and of the 1.5 Å structure of the *Malassezia sympodialis* Mala s 6 allergen, a member of the cyclophilin pan-allergen family. *Biochem J* 2006;**396**:41–49.
- Limacher A, Glaser AG, Meier C, Schmid-Grendelmeier P, Zeller S, Scapozza L et al. Cross-reactivity and 1.4-Å crystal structure of *Malassezia sympodialis* thioredoxin (Mala s 13), a member of a new pan-allergen family. *J Immunol* 2007;**178**:389–396.
- Limacher A, Kloer DP, Flückiger S, Folkers G, Crameri R, Scapozza L. The crystal structure of *Aspergillus fumigatus* cyclophilin reveals 3D domain swapping of a central element. *Structure* 2006;14:185–195.
- 78. Vilhelmsson M, Zargari A, Crameri R, Rasool O, Achour A, Scheynius A et al. Crystal structure of the major *Malassezia* sympodialis allergen Mala s 1 reveals a beta-propeller fold: a novel fold among allergens. J Mol Biol 2007;**369**:1079–1086.
- Yang X, Moffat K. Insights into specificity of cleavage and mechanisms of cell entry from the crystal structure of the highly specific Aspergillus ribotoxin, restrictocin. *Structure* 1996;4:837–852.
- Nüss D, Goettig P, Magler I, Denk U, Breitenbach M, Schneider PB et al. Crystal structure of NADP-dependent mannitol dehydrogenase from *Cladosporium herbarum*: implication for oligomerisation and catalysis. *Biochemie* 2010;92:985–993.

- Chruszcz M, Chapman MD, Osinski T, Solberg R, Demas M, Porebski PJ et al. *Alternaria alternata* allergen Alt a 1: a unique β-barrel protein dimer found exclusively in fungi. J Allergy Clin Immunol 2012;130:241–247.
- Falsone SF, Weichel M, Crameri R, Breitenbach M, Kungl AJ. Unfolding and double-stranded DNA binding of the cold shock protein homologue Cla h 8 from *Cladosporium herbarum. J Biol Chem* 2001;277:16512–16516.
- Weichel M, Schmid-Grendelmeier P, Flückiger S, Breitenbach M, Blaser K, Crameri R. Nuclear transport factor 2 represents a novel cross-reactive fungal allergen. *Allergy* 2003;58:198–206.
- 84. Rid R, Onder K, Hawranek T, Laimer M, Bauer JW, Holler C et al. Isolation and immunological characterization of a novel *Cladosporium herbarum* allergen structurally homologous to the alpha/beta hydrolase fold superfamily. *Mol Immunol* 2010;47:1366–1377.
- Flückiger S, Scapozza L, Mayer C, Blaser K, Folkers G, Crameri R. Immunological and structural analysis of IgE-mediated cross-reactivity between manganese superoxide dismutases. *Int Arch Allergy Immunol* 2002;**128**:292–303.
- Crameri R, Hemmann S, Ismail C, Menz G, Blaser K. Disease-specific recombinant allergens for the diagnosis of allergic bronchopulmonary aspergillosis. *Int Immunol* 1998;10:1211–1216.
- Hemmann S, Nikolaizik WH, Schöni MH, Blaser K, Crameri R. Differential IgE recognition of recombinant *Aspergillus fumigatus* allergens by cystic fibrosis patients with allergic bronchopulmonary aspergillosis or *Aspergillus* allergy. *Eur J Immunol* 1998;**28**:1155–1160.
- Kurup VP, Knutsen AP, Moss RB, Bansal NK. Specific antibodies to recombinant allergens of *Aspergillus fumigatus* in cystic fibrosis patients with ABPA. *Clin Mol Allergy* 2006;21:4–11.
- Knutsen AP, Slavin RG. Allergic bronchopulmonary aspergillosis in asthma and cystic fibrosis. *Clin Dev Immunol* 2011;
   2011:843763. doi: 10.1155/2011/843763.
- Slavin RG, Knutsen AP. Purified Aspergillus proteins: going where no one has gone before. J Lab Clin Med 1993;121:380–381.
- Crameri R. Recombinant Aspergillus fumigatus allergens: from the nucleotide sequences to clinical applications. Int Arch Allergy Immunol 1998;115:99–114.
- 92. Burbach GJ, Heinzerling LM, Edenharter G, Bachert C, Bindslev-Jensen C, Bonini S et al. GA(2)Len skin test study II: clinical relevance of inhalant allergen sensitization in Europe. *Allergy* 2009;**64**:1507–1515.

- Crameri R. The crux with a reliable in vitro and in vivo diagnosis of allergy. *Allergy* 2013;68:693–694.
- American Academy of Allergy. Asthma and Immunology (AAAAI). The use of standardized allergen extracts. J Allergy Clin Immunol 1997;99:583–586.
- 95. Slater JE, Menzies SL, Bridgewater J, Mosquera A, Zinderman CE, Ou AC et al. The US Food and Drug Administration review of the safety and effectiveness of nonstandardized allergen extracts. J Allergy Clin Immunol 2012;129:1014–1019.
- Wallenbeck I, Aukrust L, Einarsson R. Antigenic variability of different strains of Aspergillus fumigatus. Int Arch Allergy Appl Immunol 1984;73:166–172.
- Reed C. Variability of antigenicity of Aspergillus fumigatus. J Allergy Clin Immunol 1978;61:227–229.
- Kauffman HF, van der Heide S, van der Laan S, Hovenga H, Beaumont F, de Vries K. Standardization of allergenic extracts of *Aspergillus fumigatus*: liberation of IgEbinding components during cultivation. *Int Arch Allergy Appl Immunol* 1985;**76**:168– 173.
- Esch RE. Role of proteases on the stability of allergenic extracts. In: Klein R editor. *Regulatory Control and Standardization of Allergenic Extracts.* Stuttgart: Fischer, 1990:171–177.
- 100. Kauffman HF, van der Heide S, de Vries K. Antigenic composition of Aspergillus fumigatus in relation to the conditions of growth. In: Poucard T, Dreborg S editors. Mould Allergy Workshop. Uppsala: Pharmacia, 1994:43–44.
- Kurup VP, Shen HD, Vijay H. Immunobiology of fungal allergens. Int Arch Allergy Immunol 2002;129:181–188.
- 102. Nikolaizic WH, Crameri R, Blaser K, Schöni MH. Skin test reactivity to recombinant Aspergillus fumigatus allergen I/a in patients with cystic fibrosis. Int Arch Allergy Immunol 1996;111:403–408.
- 103. Lee JH, Park KH, Kim HS, Kim KW, Sohn MH, Kim CH et al. Specific IgE measurement using AdvanSure<sup>®</sup> system: comparison of detection performance with ImmunoCAP<sup>®</sup> system in Korean allergy patients. *Clin Chim Acta* 2012;**413**: 914–919.
- Altmann F. The role of glycoproteins in allergy. Int Arch Allergy Immunol 2007:142:99–115.
- Schmid-Grendelmeier P, Crameri R. Recombinant allergens for skin testing. Int Arch Allergy Immunol 2001;125:96–111.
- Larenas-Linnemann D, Cox LS. European allergen extract units and potency: review of available information. *Ann Allergy Asthma Immunol* 2008;100:137–145.

- 107. Hiller R, Laffer S, Harwanegg C, Huber M, Schmidt WM, Twardosz A et al. Microarrayed allergen molecules: diagnostic gatekeepers for allergy treatment. *FASEB J* 2002;16:414–416.
- Harwanegg Ch, Hiller R. Protein microarrays for the diagnosis of allergic diseases: state-of-the-art and future development. *Eur Ann Allergy Clin Immunol* 2006;**38**:232– 236.
- Lucas LM. Microarrays: molecular Allergology and nanotechnology for personalised medicine (I). *Allergol Immunopathol* (*Madr*) 2010;38:153–161.
- 110. Bonini M, Marcomini L, Gramiccioni C, Tranquilli C, Melioli G, Canonica GW et al. Microarray evaluation of specific IgE to allergen components in elite athletes. *Allergy* 2012;67:1557–1564.
- 111. Müller U, Fricker M, Wymann D, Blaser K, Crameri R. Increased specificity of diagnostic tests with recombinant major bee venom allergen phospholipase A2. *Clin Exp Allergy* 1997;27:915–920.
- Thomas WR. The advent of recombinant allergens and allergen cloning. J Allergy Clin Immunol 2011;127:855–859.
- 113. Thia LP, Balfour Lynn IM. Diagnosing allergic bronchopulmonary aspergillosis in children with cystic fibrosis. *Paediatr Respir Rev* 2009;10:37–42.
- 114. Caubet JC, Kondo Y, Urisu A, Nowak-Wegrzyn A. Molecular Diagnosis of egg allergy. *Curr Opin Allergy Clin Immunol* 2011;11:210–215.
- 115. De Knop KJ, Bridts CH, Verweij MM, Hagendorens MM, De Clerck LS, Stevens WJ et al. Component-resolved allergy diagnosis by microarrays: potential, pitfalls and prospects. *Adv Clin Chem* 2010;**50**:87–101.
- Treudler R, Simon JC. Overview of component resolved diagnostics. *Curr Allergy Asthma Rep* 2013;13:110–117.
- 117. Burr ML, Matthews IP, Arthur RA, Watson HL, Gregory CJ, Dunstan FD et al. Effects on patients with asthma of eradicating visible indoor mould: a randomized controlled trial. *Thorax* 2007;62:767–772.
- Kopp VP. Role of immunomodulators in allergen-specific immunotherapy. *Allergy* 2011;66:792–797.
- Ring J, Gutermuth J. 100 years of hyposensitization: history of allergen-specific immunotherapy (ASIT). *Allergy* 2011;66:713–724.
- Helbling A, Reimers A. Immunotherapy in fungal allergy. *Curr Allergy Asthma Rep* 2003;3:447–453.
- Malling HJ. Immunotherapy for mold allergy. Clin Rev Allergy 1992;10:237–251.
- Louis R, Schleich F, Barnes PJ. Corticosteroids: still at the frontline in asthma treatment? *Clin Chest Med* 2012;33:531–541.

- 123. Rosenberg M, Patterson R, Roberts M, Wang J. The assessment of immunologic and clinical changes occurring during corticosteroid therapy for allergic bronchopulmonary aspergillosis. *Am J Med* 1978;64:599–606.
- 124. DuBuske LM. Twenty-four-hour duration of effect of intranasal corticosteroids for seasonal allergic rhinitis symptoms: clinical evidence and relevance. *Am J Rhinol Allergy* 2012;**26**:287–292.
- 125. Seaton A, Seaton RA, Withtman AJ. Management of allergic bronchopulmonary aspergillosis without maintenance oral corticosteroids: a fifteen-year follow up. *QJM* 1994;87:529–537.
- Hilton AM, Chatterjee SS. Bronchopulmonary aspergillosis – treatment with beclomethasone dipropionate. *Postgrad Med J* 1975;**51**(Suppl 4):98–103.
- 127. Bodor N, Buchwald P. Corticosteroid design for the treatment of asthma: structural insights and the therapeutic potential of soft corticosteroids. *Curr Pharmacol Des* 2006;**12**:3241–3260.
- Wark P, Gibson PG, Wilson A. Azoles for allergic bronchopulmonary aspergillosis associated with asthma. *Cochrane Database Syst Rev* 2004;3:CD001108.
- 129. Stevens DA, Schwartz HJ, Lee JY, Moskovitz BL, Jerome DC, Catanzaro A et al. A randomized trial of itraconazole in allergic bronchopulmonary aspergillosis. *New Engl J Med* 2000;11:756–762.
- 130. Wark PA, Hensley MJ, Saltos N, Boyle MJ, Toneguzzi RC, Epid GD et al. Antiinflammatory effects of itraconazole in stable allergic bronchopulmonary aspergillosis: a randomized controlled trial. J Allergy Clin Immunol 2003;111:952–957.
- Crameri R, Blaser K. Allergy and immunity to fungal infections and colonization. *Eur Respir J* 2002;19:151–157.
- Tuite NL, Lacey K. Overview of invasive fungal infections. *Methods Mol Biol* 2013;968:1–23.
- Livermore J, Hope W. Evaluation of the pharmacokinetics and clinical utility of isavuconazole for treatment of invasive fungal infections. *Expert Opin Drug Metab Toxicol* 2012;8:759–765.
- 134. Hawranek T. Cutaneous Mycology. *Chem Immunol* 2002;**81**:129–166.
- Fishwick D. New occupational and environmental causes of asthma and extrinsic allergic alveolitis. *Clin Chest Med* 2012;33:605–616.
- Selman M, Lacasse Y, Pardo A, Cormier Y. Hypersensitivity pneumonitis caused by fungi. Proc Am Thorac Soc 2010;7:229–236.
- Kurup VP, Kumar A. Immunodiagnosis of aspergillosis. *Clin Microbiol Rev* 1991;4:439–456.

- 138. Radauer C, Nandy A, Ferreira F, Goodman RE, Larsen JN, Lidholm J et al. Update of the WHO/IUIS allergen nomenclature database based on analysis of allergen sequences. *Allergy* 2013 (in press).
- Nolles G, Hoekstra MO, Schouten JP, Gerritsen J, Kauffman HF. Prevalence of immunoglobulin E for fungi in atopic children. *Clin Exp Allergy* 2011;31:1564–1570.
- 140. Gioulekas D, Damialis A, Papakosta D, Spieksma F, Giouleka P, Patakas D. Allergenic fungi spore records (15 years) and sensitization in patients with respiratory allergy in Thessaloniki-Greece. J Investig Allereol Clin Immunol 2004:14:225–231.
- 141. Ezeamuzie CI, Al-Ali S, Khan M, Hijazi Z, Dowaisan A, Thomson MS et al. IgE-mediated sensitization to mould allergens among patients with allergic respiratory diseases in a desert environment. *Int Arch Allergy Immunol* 2000;**121**:300–307.
- 142. Green BJ, Blachere FM, Beezhold DH, Weissman DN, Hogan MB, Wilson NW et al. IgE reactivity to *Paecilomyces variotii*

antigens in fungal sensitized patients. J Allergy Clin Immunol 2007;119:S187.

- Simons FE, Sheikh A. Anaphylaxis: the acute episode and beyond. *BMJ* 2013;346: f602.
- 144. Kalish RS, Askenase PW. Molecular mechanisms of CD8 + T cell-mediated delayed hypersensitivity: implications for allergies, asthma, and autoimmunity. J Allergy Clin Immunol 1999;103:192–199.
- 145. Karsten CM, Köhl J. The immunoglobulin, IgG Fc receptor and complement triangle in autoimmune diseases. *Immunobiol* 2012;217:1067–1079.
- Spiewak R. Contact dermatitis in atopic individuals. Curr Opin Allergy Clin Immunol 2012;12:491–497.
- 147. Gideon HP, Flynn JL. Latent tuberculosis: what the host "sees"? *Immunol Res* 2011;50:202–212.
- 148. Moser M, Crameri R, Menz G, Schneider T, Dudler T, Virchow C et al. Cloning and expression of recombinant *Aspergillus fumigatus* allergenI/a (rAsp f I/a) with IgE

binding and type I skin test activity. J Immunol 1992;**149**:454–460.

- Flückiger S, Fijten H, Whitley P, Blaser K, Crameri R. Cyclophilins, a new family of cross-reactive allergens. *Eur J Immunol* 2002;32:10–17.
- 150. Glaser AG, Kirsch AI, Zeller S, Menz G, Rhyner C, Crameri R. Molecular and immunological characterization of Asp f 34, a novel major cell wall allergen of *Aspergilus fumigatus. Allergy* 2009;64:1144– 1151.
- 151. Schneider PB, Denk U, Breitenbach M, Richter K, Schmid-Grendelmeier P, Nobbe S et al. *Alternaria alternata* NADPdependent mannitol dehydrogenase is an important fungal allergen. *Clin Exp Allergy* 2006;**36**:1513–1524.
- 152. Shen HD, Tam MF, Chou H, Han SH. The importance of serine proteinases as aeroallergens associated with asthma. Int Arch Allergy Immunol 1999;119:259–264.